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(54) Title: CHEMOKINE ALPHA-5 (57) Abstract The present invention relates to a novel CK α -5 protein which is a member of the alpha chemokine family. In particular, isolated nucleic acid molecules are provided encoding the human CK α -5 protein. CK α -5 polypeptides are also provided as are vectors, host cells and recombinant methods for producing the same. The invention further relates to screening methods for identifying agonists and antagonists of CK α -5 activity. Also provided are diagnostic methods for detecting immune system-related disorders and therapeutic methods for treating immune system-related disorders.		

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Chemokine Alpha-5

Field of the Invention

The present invention relates to a novel human gene encoding a polypeptide which is a member of the chemokine family. More specifically, isolated nucleic acid molecules are provided encoding a human polypeptide named Chemokine Alpha-5, hereinafter referred to as "CK α -5".

CK α -5 polypeptides are also provided, as are vectors, host cells and recombinant methods for producing the same. Also provided are diagnostic methods for detecting disorders related to the immune system, and therapeutic methods for treating such disorders. The invention further relates to screening methods for identifying agonists and antagonists of CK α -5 activity.

Background of the Invention

The ability to control the migration and "trafficking" of various cell types is controlled by a subset of factors, or proteins, among which chemokines are an example.

Chemokines, also referred to as intercrine cytokines, are a subfamily of structurally and functionally related chemotactic cytokines. These molecules are usually 8-10 kd in size, however, larger membrane bound chemokine polypeptides have been identified. In general, chemokines exhibit 20% to 75% homology at the amino acid level and are characterized by four conserved cysteine residues that form two disulfide bonds. Based on the arrangement of the first two cysteine residues, chemokines have been classified into two subfamilies, alpha and beta. In the alpha subfamily, the first two cysteines are separated by one amino acid and hence are referred to as the "C-X-C" subfamily. In the beta subfamily, the two cysteines are in an adjacent position and are, therefore, referred to as the "C-C" subfamily. Thus far, at least sixteen different members of this family have been identified in humans.

The intercrine cytokines exhibit a wide variety of functions. A hallmark feature is their ability to elicit chemotactic migration of distinct cell types, including monocytes, neutrophils, T lymphocytes, basophils and fibroblasts. Many chemokines have proinflammatory activity and are involved in multiple steps during an inflammatory reaction. These activities include stimulation of histamine release, lysosomal enzyme and leukotriene release, increased adherence of target immune cells to endothelial cells, enhanced binding of complement proteins, induced expression of granulocyte adhesion molecules and complement receptors, and respiratory burst. In addition to their involvement in inflammation, certain chemokines have been shown to exhibit other activities. For example, macrophage inflammatory protein 1 (MIP-1) is able to suppress hematopoietic stem cell proliferation, platelet factor-4 (PF-4) is a potent inhibitor of endothelial cell growth, Interleukin-8 (IL-8) promotes proliferation of keratinocytes, and GRO is an autocrine growth factor for melanoma cells.

In light of the diverse biological activities, it is not surprising that chemokines have been implicated in a number of physiological and disease conditions, including lymphocyte trafficking, wound healing, hematopoietic regulation and immunological disorders such as allergy, asthma and arthritis.

5 Members of the "C-C" branch exert their effects on the following cells: eosinophils which destroy parasites to lessen parasitic infection and cause chronic inflammation in the airways of the respiratory system; macrophages which suppress tumor formation in vertebrates; and basophils which release histamine which plays a role in allergic inflammation. However, members of one branch may exert an effect on cells which are normally responsive to the other branch of
10 chemokines and, therefore, no precise role can be attached to the members of the branches.

While members of the C-C branch act predominantly on mononuclear cells and members of the C-X-C branch act predominantly on neutrophils a distinct chemoattractant property cannot be assigned to a chemokine based on this guideline. Some chemokines from one family show characteristics of the other.

15 The polypeptide of the present invention has the conserved cysteine residues of the "C-X-C" region, and has amino acid sequence homology to known chemokines.

Thus, there is a need for polypeptides that function as regulators of the migration of distinct cell types and of their roles in dysfunction and disease, since disturbances of such regulation may be involved in disorders relating to hemostasis, angiogenesis, tumor metastasis,
20 cellular migration and ovulation, as well as neurogenesis. Therefore, there is a need for identification and characterization of such human polypeptides which can play a role in detecting, preventing, ameliorating or correcting such disorders.

Summary of the Invention

The present invention provides isolated nucleic acid molecules comprising a
25 polynucleotide encoding at least a portion of the CK α -5 polypeptide having the complete amino acid sequence shown in SEQ ID NO:2 or the complete amino acid sequence encoded by a cDNA clone deposited as plasmid DNA as ATCC Deposit Number 209231 on August 29, 1997. The ATCC is located at 10801 University Boulevard, Manassas, Virginia 20110-2209. The nucleotide sequence determined by sequencing the deposited CK α -5 clone, which is shown in Figure 1
30 (SEQ ID NO:1), contains an open reading frame encoding a complete polypeptide of 254 amino acid residues, including an initiation codon encoding an N-terminal methionine at nucleotide positions 542-544. Nucleic acid molecules of the invention include those encoding the complete amino acid sequence excepting the N-terminal methionine shown in SEQ ID NO:2, or the complete amino acid sequence excepting the N-terminal methionine encoded by a cDNA clone in
35 ATCC Deposit Number 209231, which molecules also can encode additional amino acids fused to the N-terminus of the CK α -5 amino acid sequence.

The polypeptide of the present invention has amino acid sequence homology to known chemokines, including the conserved C-X-C cysteine pattern characteristic of the alpha subfamily of chemokines beginning with the first cysteine from the amino terminus in SEQ ID NO:2.

CK α -5 also lacks the ELR motif found in some alpha chemokines immediately preceding
5 the first cysteine residue, which is known to be required for the neutrophil and endothelial cell chemotactic activity as well as the angiogenic activity of IL-8.

The encoded polypeptide has a predicted leader sequence of 27 amino acids underlined in Figure 1; and the amino acid sequence of the predicted mature CK α -5 protein is also shown in Figure 1 as amino acid residues 28-254 and as residues 1-227 in SEQ ID NO:2.
10 Further, the polypeptide is predicted to consist of an extracellular domain having the sequence from residues 28 to 205 in Figure 1 (1 to 178 in SEQ ID NO:2), a transmembrane domain having a sequence from residue 206 to 227 in Figure 1 (179 to 200 in SEQ ID NO:2), and an intracellular domain having a sequence from residue 228 to 254 in Figure 1 (201 to 227 in SEQ ID NO:2).

Thus, one aspect of the invention provides an isolated polynucleotide comprising a
15 nucleotide sequence selected from the group consisting of: (a) a nucleotide sequence encoding the CK α -5 polypeptide having the complete amino acid sequence in Figure 1 (SEQ ID NO:2); (b) a nucleotide sequence encoding the CK α -5 polypeptide having the complete amino acid sequence in Figure 1 (SEQ ID NO:2) excepting the N-terminal methionine (i.e., positions -26 to 227 of SEQ ID NO:2); (c) a nucleotide sequence encoding the predicted mature CK α -5 polypeptide having
20 the amino acid sequence at positions 28-254 in Figure 1 (1-227 in SEQ ID NO:2); (d) a nucleotide sequence encoding the predicted extracellular domain of the CK α -5 polypeptide having the amino acid sequence at positions 28-205 in Figure 1 (1-178 in SEQ ID NO:2); (e) a nucleotide sequence encoding the CK α -5 polypeptide having the complete amino acid sequence encoded by the cDNA clone contained in ATCC Deposit No. 209231; (f) a nucleotide sequence
25 encoding the CK α -5 polypeptide having the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA clone contained in ATCC Deposit No. 209231; (g) a nucleotide sequence encoding the mature CK α -5 polypeptide encoded by the cDNA clone contained in ATCC Deposit No. 209231; (h) a nucleotide sequence encoding the extracellular domain of CK α -5 encoded by the cDNA clone contained in ATCC Deposit No. 209231; and (i) a
30 nucleotide sequence complementary to any of the nucleotide sequences in (a), (b), (c), (d), (e), (f), (g) or (h) above.

Further embodiments of the invention include isolated nucleic acid molecules that comprise a polynucleotide having a nucleotide sequence at least 90% identical, and more preferably at least 95%, 96%, 97%, 98% or 99% identical, to any of the nucleotide sequences in
35 (a), (b), (c), (d), (e), (f), (g), (h), or (i) above, or a polynucleotide which hybridizes under

stringent hybridization conditions to a polynucleotide in (a), (b), (c), (d), (e), (f), (g), (h) or (i), above. This polynucleotide which hybridizes does not hybridize under stringent hybridization conditions to a polynucleotide having a nucleotide sequence consisting of only A residues or of only T residues. An additional nucleic acid embodiment of the invention relates to an isolated
5 nucleic acid molecule comprising a polynucleotide which encodes the amino acid sequence of an epitope-bearing portion of a CK α -5 polypeptide having an amino acid sequence in (a), (b), (c), (d), (e), (f), (g) or (h), above.

The present invention also relates to recombinant vectors, which include the isolated nucleic acid molecules of the present invention, and to host cells containing the recombinant
10 vectors, as well as to methods of making such vectors and host cells and for using them for production of CK α -5 polypeptides or peptides by recombinant techniques.

The invention further provides an isolated CK α -5 polypeptide comprising an amino acid sequence selected from the group consisting of: (a) the amino acid sequence of the full-length CK α -5 polypeptide having the complete amino acid sequence shown in SEQ ID NO:2 or the
15 complete amino acid sequence encoded by the cDNA clone contained in the ATCC Deposit No. 209231; (b) the amino acid sequence of the full-length CK α -5 polypeptide having the complete amino acid sequence shown in SEQ ID NO:2 excepting the N-terminal methionine (i.e., positions -26 to 227 of SEQ ID NO:2) or the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA clone contained in the ATCC Deposit No. 209231; (c) the
20 amino acid sequence of the mature CK α -5 polypeptide having the amino acid sequence shown in SEQ ID NO:2 as residues 1-227 or the amino acid sequence of the mature CK α -5 encoded by the cDNA clone contained in the ATCC Deposit No. 209231; and (d) the amino acid sequence of the extracellular domain of the CK α -5 polypeptide having the amino acid sequence shown in SEQ ID NO:2 as residues 1-178 or the amino acid sequence of the extracellular domain of CK α -5
25 encoded by the cDNA clone contained in the ATCC Deposit No. 209231. The polypeptides of the present invention also include polypeptides having an amino acid sequence at least 80% identical, more preferably at least 90% identical, and still more preferably 95%, 96%, 97%, 98% or 99% identical to those described in (a), (b), (c) or (d) above, as well as polypeptides having an amino acid sequence with at least 90% similarity, and more preferably at least 95% similarity, to
30 those above. Polynucleotides encoding polypeptides having an amino acid sequence at least 80% identical, more preferably at least 90% identical, and still more preferably 95%, 96%, 97%, 98% or 99% identical to those described in (a), (b), (c) or (d) above, as well as polypeptides having an amino acid sequence with at least 90% similarity, and more preferably at least 95% similarity, to those above are also provided.

An additional embodiment of this aspect of the invention relates to a peptide or polypeptide which comprises the amino acid sequence of an epitope-bearing portion of a CK α -5 polypeptide having an amino acid sequence described in (a), (b), (c) or (d), above. Peptides or polypeptides having the amino acid sequence of an epitope-bearing portion of a CK α -5

5 polypeptide of the invention include portions of such polypeptides with at least six (6) or seven (7), preferably at least nine (9), and more preferably at least about 30 amino acids to about 50 amino acids, although epitope-bearing polypeptides of any length up to and including the entire amino acid sequence of a polypeptide of the invention described above also are included in the invention.

10 In another embodiment, the invention provides an isolated antibody that binds specifically to a CK α -5 polypeptide having an amino acid sequence described in (a), (b), (c) or (d) above.

The invention further provides methods for isolating antibodies that bind specifically to a CK α -5 polypeptide having an amino acid sequence as described herein. Such antibodies are useful diagnostically or therapeutically as described below.

15 The invention also provides for pharmaceutical compositions comprising CK α -5 polypeptides, particularly human CK α -5 polypeptides, which may be employed, for instance, to stimulate wound healing, to treat solid tumors, microbial infections, autoimmune diseases, liver cirrhosis, osteoarthritis and pulmonary fibrosis. Methods of treating individuals in need of CK α -5 polypeptides are also provided.

20 The invention further provides compositions comprising a CK α -5 polynucleotide or a CK α -5 polypeptide for administration to cells *in vitro*, to cells *ex vivo* and to cells *in vivo*, or to a multicellular organism. In certain particularly preferred embodiments of this aspect of the invention, the compositions comprise a CK α -5 polynucleotide for expression of a CK α -5 polypeptide in a host organism for treatment of disease. Particularly preferred in this regard is
25 expression in a human patient for treatment of a dysfunction associated with aberrant endogenous activity of a CK α -5

The present invention also provides a screening method for identifying compounds capable of enhancing or inhibiting a biological activity of the CK α -5 polypeptide, which involves contacting a receptor which is enhanced by the CK α -5 polypeptide with the candidate compound
30 in the presence of a CK α -5 polypeptide, assaying calcium mobilization or chemotactic activity of the cell expressing the receptor in the presence of the candidate compound and the CK α -5 polypeptide, and comparing the receptor activity to a standard level of activity, the standard being assayed when contact is made between the receptor and the CK α -5 polypeptide in the absence of

the candidate compound. In this assay, an increase in calcium mobilization or chemotaxis over the standard indicates that the candidate compound is an agonist of CK α -5 activity and a decrease in calcium mobilization or chemotaxis compared to the standard indicates that the compound is an antagonist of CK α -5 activity.

It has been discovered that CK α -5 is expressed not only in neutrophils but also in monocytes, macrophages, liver, lung, testes, cerebellum and pineal gland. Therefore, nucleic acids of the invention are useful as hybridization probes for differential identification of the tissue(s) or cell type(s) present in a biological sample. Similarly, polypeptides and antibodies directed to those polypeptides are useful to provide immunological probes for differential identification of the tissue(s) or cell type(s). In addition, for a number of disorders of the above tissues or cells, particularly of the immune system, significantly higher or lower levels of CK α -5 gene expression may be detected in certain tissues (e.g., cancerous and wounded tissues) or bodily fluids (e.g., serum, plasma, urine, synovial fluid or spinal fluid) taken from an individual having such a disorder, relative to a "standard" CK α -5 gene expression level, i.e., the CK α -5 expression level in healthy tissue from an individual not having the immune system disorder. Thus, the invention provides a diagnostic method useful during diagnosis of such a disorder, which involves: (a) assaying CK α -5 gene expression level in cells or body fluid of an individual; (b) comparing the CK α -5 gene expression level with a standard CK α -5 gene expression level, whereby an increase or decrease in the assayed CK α -5 gene expression level compared to the standard expression level is indicative of disorder in the immune system.

An additional aspect of the invention is related to a method for treating an individual in need of an increased level of CK α -5 activity in the body comprising administering to such an individual a composition comprising a therapeutically effective amount of an isolated CK α -5 polypeptide of the invention or an agonist thereof.

A still further aspect of the invention is related to a method for treating an individual in need of a decreased level of CK α -5 activity in the body comprising, administering to such an individual a composition comprising a therapeutically effective amount of a CK α -5 antagonist. Preferred antagonists for use in the present invention are CK α -5-specific antibodies.

Brief Description of the Figures

Figure 1A-C shows the nucleotide sequence (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO:2) of CK α -5. The predicted leader sequence of about 27 amino acids is underlined. Note that the methionine residue at the beginning of the leader sequence in Figure 1

is shown in position number (positive) 1, whereas the leader positions in the corresponding sequence of SEQ ID NO:2 are designated with negative position numbers. Thus, the leader sequence positions 1 to 27 in Figure 1 correspond to positions -27 to -1 in SEQ ID NO:2.

Figure 2 shows the regions of identity between the amino acid sequence of the CK α -5 protein and translation product of the rat mRNA for MIP1- β (SEQ ID NO:3), as determined by Bestfit (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711) using the default parameters.

Figure 3 shows an analysis of the CK α -5 amino acid sequence. Alpha, beta, turn and coil regions; hydrophilicity and hydrophobicity; amphipathic regions; flexible regions; antigenic index and surface probability are shown. In the "Antigenic Index - Jameson-Wolf" graph, the positive peaks indicate locations of the highly antigenic regions of the CK α -5 protein, i.e., regions from which epitope-bearing peptides of the invention can be obtained.

Detailed Description

The present invention provides isolated nucleic acid molecules comprising a polynucleotide encoding a CK α -5 polypeptide having the amino acid sequence shown in SEQ ID NO:2, which was determined by sequencing cloned cDNAs. The nucleotide sequence shown in Figure 1 (SEQ ID NO:1) was obtained by sequencing two cDNA clones: HMSCJ62 and HNFEM05. HMSCJ62 contains the full-length sequence shown in Figure 1 and is deposited at Human Genome Sciences, Inc., 9410 Key West Ave., Rockville, MD 20850. HNFEM05 contains all but the first 360 nucleotides (including all of the coding region) and was deposited as American Type Culture Collection ("ATCC") Deposit No. 209231 on August 29, 1997. The ATCC is located at 10801 University Boulevard, Manassas, Virginia 20110-2209.

The deposited clones are contained in the pBluescript SK(-) plasmid (Stratagene, La Jolla, CA).

The polypeptide of the present invention has amino acid sequence homology to known chemokines, including the conserved C-X-C cysteine pattern characteristic of the alpha subfamily of chemokines beginning with the first cysteine from the amino terminus in SEQ ID NO:2. The CK α -5 protein of the present invention also shares sequence homology with other chemokines including in particular the translation product of the human mRNA for MIP1- β (SEQ ID NO:3) as shown in Figure 2.

Of the known members of the alpha chemokine family, the majority contain an ELR motif (e.g., IL-8, ENA-78, GCP2, GRO- α , PBP, CTAP-III and NAP-2) and others lack the ELR motif (IP-10, PF4 and MIG). CK α -5 lacks the ELR motif immediately preceding the first cysteine residue. It has been clearly shown that this ELR motif is required for the neutrophil and

endothelial cell chemotactic activity as well as the angiogenic activity of IL-8 (Strieter *et al.*, *J. Biol. Chem.*, 270:27348-27357 (1995)). In addition, it has been shown that the ELR-C-X-C chemokines have *in vitro* and *in vivo* angiogenic activity, whereas the C-X-C chemokines lacking the ELR motif are actually angiostatic (Strieter *et al.*, *supra*; Angiolillo *et al.*, *J. Exp. Med.*, 182:155-162 (1995); and Koch *et al.*, *Science*, 258:1798-1801 (1992)). In terms of a possible role of such factors in tumor angiogenesis, Smith *et al.*, *J. Exp. Med.*, 179:1409-1415 (1994), has reported increased IL-8 levels in bronchogenic carcinoma tumor tissues which appear to be produced from the tumor cells.

Nucleic Acid Molecules

Unless otherwise indicated, all nucleotide sequences determined by sequencing a DNA molecule herein were determined using an automated DNA sequencer (such as the Model 373 from Applied Biosystems, Inc., Foster City, CA), and all amino acid sequences of polypeptides encoded by DNA molecules determined herein were predicted by translation of a DNA sequence determined as above. Therefore, as is known in the art for any DNA sequence determined by this automated approach, any nucleotide sequence determined herein may contain some errors. Nucleotide sequences determined by automation are typically at least about 90% identical, more typically at least about 95% to at least about 99.9% identical to the actual nucleotide sequence of the sequenced DNA molecule. The actual sequence can be more precisely determined by other approaches including manual DNA sequencing methods well known in the art. As is also known in the art, a single insertion or deletion in a determined nucleotide sequence compared to the actual sequence will cause a frame shift in translation of the nucleotide sequence such that the predicted amino acid sequence encoded by a determined nucleotide sequence will be completely different from the amino acid sequence actually encoded by the sequenced DNA molecule, beginning at the point of such an insertion or deletion.

By "nucleotide sequence" of a nucleic acid molecule or polynucleotide is intended, for a DNA molecule or polynucleotide, a sequence of deoxyribonucleotides, and for an RNA molecule or polynucleotide, the corresponding sequence of ribonucleotides (A, G, C and U), where each thymidine deoxyribonucleotide (T) in the specified deoxyribonucleotide sequence is replaced by the ribonucleotide uridine (U).

Using the information provided herein, such as the nucleotide sequence in Figure 1 (SEQ ID NO:1), a nucleic acid molecule of the present invention encoding a CK α -5 polypeptide may be obtained using standard cloning and screening procedures, such as those for cloning cDNAs using mRNA as starting material. Illustrative of the invention, the nucleic acid molecule described in Figure 1 (SEQ ID NO:1) was discovered in cDNA libraries derived primarily from immune-system tissues. Clone HMSCJ62 was isolated from a monocyte cDNA library and HNFEM05 was isolated from a neutrophil cDNA library.

Additional clones of the same gene were also identified in cDNA libraries from the following human tissues: colon and osteoclastoma.

The determined nucleotide sequence of the CK α -5 cDNA of Figure 1 (SEQ ID NO:1) contains an open reading frame encoding a protein of 254 amino acid residues, with an initiation codon at nucleotide positions 542-544 of the nucleotide sequence in Figure 1 (SEQ ID NO:1). The amino acid sequence of the CK α -5 protein shown in SEQ ID NO:2 is about 51% similar and
5 28 % identical to the rat Macrophage Inflammatory Protein-1 β (Figure 2), which can be accessed on GenBank as Accession No. gil459148.

As one of ordinary skill would appreciate, due to the possibilities of sequencing errors discussed above, the actual complete CK α -5 polypeptide encoded by the deposited cDNA, which comprises about 254 amino acids, may be somewhat longer or shorter. More generally, the
10 actual open reading frame may be anywhere in the range of ± 20 amino acids, more likely in the range of ± 10 amino acids, of that predicted from the methionine codon at the N-terminus shown in Figure 1 (SEQ ID NO:1).

Leader and Mature Sequences

The amino acid sequence of the complete CK α -5 protein includes a leader sequence and a
15 mature protein, as shown in SEQ ID NO:2. More in particular, the present invention provides nucleic acid molecules encoding a mature form of the CK α -5 protein. Thus, according to the signal hypothesis, once export of the growing protein chain across the rough endoplasmic reticulum has been initiated, proteins secreted by mammalian cells have a signal or secretory leader sequence which is cleaved from the complete polypeptide to produce a secreted "mature"
20 form of the protein. Most mammalian cells and even insect cells cleave secreted proteins with the same specificity. However, in some cases, cleavage of a secreted protein is not entirely uniform, which results in two or more mature species of the protein. Further, it has long been known that the cleavage specificity of a secreted protein is ultimately determined by the primary structure of the complete protein, that is, it is inherent in the amino acid sequence of the polypeptide.
25 Therefore, the present invention provides a nucleotide sequence encoding the mature CK α -5 polypeptide having the amino acid sequence encoded by the cDNA clone identified as ATCC Deposit No. 209231. By the "mature CK α -5 polypeptide having the amino acid sequence encoded by the cDNA clone in ATCC Deposit No. 209231" is meant the mature form(s) of the CK α -5 protein produced by expression in a mammalian cell (e.g., COS cells, as described
30 below) of the complete open reading frame encoded by the human DNA sequence of the clone contained in the deposited vector.

In addition, methods for predicting whether a protein has a secretory leader as well as the cleavage point for that leader sequence are available. For instance, the method of McGeoch (*Virus Res.* 3:271-286 (1985)) uses the information from a short N-terminal charged region and a

subsequent uncharged region of the complete (uncleaved) protein. The method of von Heinje (*Nucleic Acids Res.* 14:4683-4690 (1986)) uses the information from the residues surrounding the cleavage site, typically residues -13 to +2 where +1 indicates the amino terminus of the mature protein. The accuracy of predicting the cleavage points of known mammalian secretory proteins for each of these methods is in the range of 75-80% (von Heinje, *supra*). However, the two methods do not always produce the same predicted cleavage point(s) for a given protein.

In the present case, the deduced amino acid sequence of the complete CK α -5 polypeptide was analyzed by a computer program "PSORT", available from Dr. Kenta Nakai of the Institute for Chemical Research, Kyoto University (see K. Nakai and M. Kanehisa, *Genomics* 14:897-911 (1992)), which is an expert system for predicting the cellular location of a protein based on the amino acid sequence. As part of this computational prediction of localization, the methods of McGeoch and von Heinje are incorporated." The computation analysis above predicted one potential cleavage site within the complete amino acid sequence shown in SEQ ID NO:2; that is, between residues 27 and 28 in Figure 1 and residues -1 and 1 in SEQ ID NO:2.

As indicated, nucleic acid molecules of the present invention may be in the form of RNA, such as mRNA, or in the form of DNA, including, for instance, cDNA and genomic DNA obtained by cloning or produced synthetically. The DNA may be double-stranded or single-stranded. Single-stranded DNA or RNA may be the coding strand, also known as the sense strand, or it may be the non-coding strand, also referred to as the anti-sense strand.

By "isolated" nucleic acid molecule(s) is intended a nucleic acid molecule, DNA or RNA, which has been removed from its native environment. For example, recombinant DNA molecules contained in a vector are considered isolated for the purposes of the present invention. Further examples of isolated DNA molecules include recombinant DNA molecules maintained in heterologous host cells or purified (partially or substantially) DNA molecules in solution.

Isolated RNA molecules include *in vivo* or *in vitro* RNA transcripts of the DNA molecules of the present invention. Isolated nucleic acid molecules according to the present invention further include such molecules produced synthetically. By "isolated" the inventors intend to specifically exclude very large pieces of genetic material (200 kb +) such as isolated chromosomes and yeast artificial chromosomes (YACs).

Isolated nucleic acid molecules of the present invention include DNA molecules comprising an open reading frame (ORF) with an initiation codon at positions 542-544 of the nucleotide sequence shown in Figure 1 (SEQ ID NO:1).

Also included are DNA molecules comprising the coding sequence for the predicted mature CK α -5 protein shown at positions 1-227 of SEQ ID NO:2.

In addition, isolated nucleic acid molecules of the invention include DNA molecules which comprise a sequence substantially different from those described above but which, due to the degeneracy of the genetic code, still encode a CK α -5 protein. Of course, the genetic code and species-specific codon preferences are well known in the art. Thus, it would be routine for one

skilled in the art to generate the degenerate variants described above, for instance, to optimize codon expression for a particular host (e.g., change codons in the human mRNA to those preferred by a bacterial host such as *E. coli*).

In another aspect, the invention provides isolated nucleic acid molecules encoding the CK α -5 polypeptide having an amino acid sequence encoded by the cDNA clone contained in the plasmid deposited as ATCC Deposit No. 209231 on August 29, 1997.

Preferably, this nucleic acid molecule will encode the mature polypeptide encoded by the above-described deposited cDNA clone.

The invention further provides an isolated nucleic acid molecule having the nucleotide sequence shown in Figure 1 (SEQ ID NO:1) or the nucleotide sequence of the CK α -5 cDNA contained in the above-described deposited clone, or a nucleic acid molecule having a sequence complementary to one of the above sequences. Such isolated molecules, particularly DNA molecules, are useful as probes for gene mapping, by *in situ* hybridization with chromosomes, and for detecting expression of the CK α -5 gene in human tissue, for instance, by Northern blot analysis.

The present invention is further directed to nucleic acid molecules encoding portions of the nucleotide sequences described herein as well as to fragments of the isolated nucleic acid molecules described herein. In particular, the invention provides a polynucleotide comprising a nucleotide sequence representing the portion of SEQ ID NO:1 which consists of positions 1-1303, 542-1303, 622-1303, 625-1303, 628-1303, 631-1303, 634-1303, 637-1303, 640-1303, 643-1303, 646-1303, 649-1303, 652-1303, 652-787, 652-817, 652-847, 652-877, 652-907, 652-937, 652-967, 652-997, 652-1027, 652-1057, 652-1087, 652-1117, 652-1147, 652-1177, 652-1207, 652-1237, or 652-1267 of SEQ ID NO:1.

In addition, the following nucleic acid molecules having nucleotide sequences related to extensive portions of SEQ ID NO:1 have been identified: HNFCO09R (SEQ ID NO:4) from clone HNFCO09; HNEDS26R (SEQ ID NO:5) from clone HNEDS26R; HOABD47R (SEQ ID NO:6) from clone HOABD47; HTEIR41R (SEQ ID NO:7) from clone HTEIR41; HMNAD36R (SEQ ID NO:8) from clone HMNAD36; and HTEDQ07RA (SEQ ID NO:20) from clone HTEDQ07.

The following publicly available expressed sequence tags ("ESTs"), which relate to portions of SEQ ID NO:1, have also been identified: GenBank Accession No. AA130776 (SEQ ID NO:9); GenBank Accession No. AA290712 (SEQ ID NO:10); GenBank Accession No. AA366329 (SEQ ID NO:11); GenBank Accession No. AA121716 (SEQ ID NO:12); GenBank Accession No. AA146672 (SEQ ID NO:13); GenBank Accession No. AA149359 (SEQ ID NO:14); GenBank Accession No. AA569970 (SEQ ID NO:21); and GenBank Accession No. AA994552 (SEQ ID NO:22).

Polynucleotides of the invention preferably are not, and do not comprise, one or more of the forgoing sequences 1-14 and 20-22.

Further, the invention includes a polynucleotide comprising any portion of at least about 17 nucleotides, preferably at least about 20, 25, or 30, and most preferably at least about 50 nucleotides, of SEQ ID NO:1 from nucleotide 542 to 1303.

The invention also includes a polynucleotide comprising any portion of at least about 500, preferably about 600 nucleotides, of SEQ ID NO:1, preferably from residue 542 to 1,303. Particularly preferred nucleotide fragments in this regard include the fragment from nucleotide 622 to 1,303 in SEQ ID NO:1.

More generally, by a fragment of an isolated nucleic acid molecule having the nucleotide sequence of the deposited cDNA or the nucleotide sequence shown in Figure 1 (SEQ ID NO:1) is intended fragments at least about 15 nt, and more preferably at least about 20 nt, still more preferably at least about 30 nt, and even more preferably, at least about 40 nt in length which are useful as diagnostic probes and primers as discussed herein. Of course, larger fragments 50-1,000 nt in length are also useful according to the present invention as are fragments corresponding to most, if not all, of the nucleotide sequence of the deposited cDNA or as shown in Figure 1 (SEQ ID NO:1). By a fragment at least 20 nt in length, for example, is intended fragments which include 20 or more contiguous bases from the nucleotide sequence of the deposited cDNA or the nucleotide sequence as shown in Figure 1 (SEQ ID NO:1). Preferred nucleic acid fragments of the present invention include nucleic acid molecules encoding epitope-bearing portions of the CK α -5 polypeptide as identified from the Jameson-Wolf antigenic index shown in Figure 3 and described in more detail below.

In another aspect, the invention provides an isolated nucleic acid molecule comprising a polynucleotide which hybridizes under stringent hybridization conditions to a portion of the polynucleotide in a nucleic acid molecule of the invention described above, for instance, the cDNA clone contained in ATCC Deposit No. 209231. By "stringent hybridization conditions" is intended overnight incubation at 42° C in a solution comprising: 50% formamide, 5x SSC (750 mM NaCl, 75 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 μ g/ml denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1x SSC at about 65° C.

By a polynucleotide which hybridizes to a "portion" of a polynucleotide is intended a polynucleotide (either DNA or RNA) hybridizing to at least about 15 nucleotides (nt), and more preferably at least about 20 nt, still more preferably at least about 30 nt, and even more preferably about 30-70 (e.g., 50) nt of the reference polynucleotide. These are useful as diagnostic probes and primers as discussed above and in more detail below.

By a portion of a polynucleotide of "at least 20 nt in length," for example, is intended 20 or more contiguous nucleotides from the nucleotide sequence of the reference polynucleotide (e.g., the deposited cDNA or the nucleotide sequence as shown in Figure 1 (SEQ ID NO:1)). Of course, a polynucleotide which hybridizes only to a poly A sequence (such as the 3' terminal poly(A) tract of the CK α -5 cDNA shown in Figure 1 (SEQ ID NO:1)), or to a complementary stretch of T (or U) residues, would not be included in a polynucleotide of the invention used to

hybridize to a portion of a nucleic acid of the invention, since such a polynucleotide would hybridize to any nucleic acid molecule containing a poly (A) stretch or the complement thereof (e.g., practically any double-stranded cDNA clone).

As indicated, nucleic acid molecules of the present invention which encode a CK α -5 polypeptide may include, but are not limited to those encoding the amino acid sequence of the mature polypeptide, by itself; and the coding sequence for the mature polypeptide and additional sequences, such as those encoding the about 27 amino acid leader or secretory sequence, such as a pre-, or pro- or prepro- protein sequence; the coding sequence of the mature polypeptide, with or without the aforementioned additional coding sequences.

Also encoded by nucleic acids of the invention are the above protein sequences together with additional, non-coding sequences, including for example, but not limited to introns and non-coding 5' and 3' sequences, such as the transcribed, non-translated sequences that play a role in transcription, mRNA processing, including splicing and polyadenylation signals, for example - ribosome binding and stability of mRNA; an additional coding sequence which codes for additional amino acids, such as those which provide additional functionalities.

Thus, the sequence encoding the polypeptide may be fused to a marker sequence, such as a sequence encoding a peptide which facilitates purification of the fused polypeptide. In certain preferred embodiments of this aspect of the invention, the marker amino acid sequence is a hexa-histidine peptide, such as the tag provided in a pQE vector (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311), among others, many of which are commercially available. As described in Gentz *et al.*, *Proc. Natl. Acad. Sci. USA* 86:821-824 (1989), for instance, hexa-histidine provides for convenient purification of the fusion protein. The "HA" tag is another peptide useful for purification which corresponds to an epitope derived from the influenza hemagglutinin protein, which has been described by Wilson *et al.*, *Cell* 37: 767 (1984). As discussed below, other such fusion proteins include the CK α -5 fused to Fc at the N- or C-terminus.

Variant and Mutant Polynucleotides

The present invention further relates to variants of the nucleic acid molecules of the present invention, which encode portions, analogs or derivatives of the CK α -5 protein. Variants may occur naturally, such as a natural allelic variant. By an "allelic variant" is intended one of several alternate forms of a gene occupying a given locus on a chromosome of an organism. *Genes II*, Lewin, B., ed., John Wiley & Sons, New York (1985). Non-naturally occurring variants may be produced using art-known mutagenesis techniques.

Such variants include those produced by nucleotide substitutions, deletions or additions. The substitutions, deletions or additions may involve one or more nucleotides. The variants may be altered in coding regions, non-coding regions, or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions or additions.

Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of the CK α -5 protein or portions thereof. Also especially preferred in this regard are conservative substitutions.

Most highly preferred are nucleic acid molecules encoding the mature protein having the amino acid sequence shown in SEQ ID NO:2 or the mature CK α -5 amino acid sequence encoded by the deposited cDNA clone either of which may be modified so as to encode an amino terminal methionine or other N- or C-terminal fusion peptides or polypeptides.

Further embodiments include an isolated polynucleotide comprising a nucleotide sequence at least 90% identical, and more preferably at least 95%, 96%, 97%, 98% or 99% identical to a nucleic acid sequence selected from the group consisting of: (a) a nucleotide sequence encoding the CK α -5 polypeptide shown as SEQ ID NO:2; (b) a nucleotide sequence encoding the CK α -5 polypeptide shown as residues -26 to 227 of SEQ ID NO:2; (c) a nucleotide sequence encoding the predicted mature CK α -5 shown as residues 1-227 in SEQ ID NO:2; (d) a nucleotide sequence encoding the predicted extracellular domain of the CK α -5 polypeptide shown as

residues 1-178 in SEQ ID NO:2; (e) a nucleotide sequence encoding the CK α -5 polypeptide having the complete amino acid sequence encoded by the cDNA clone contained in ATCC Deposit No. 209231; (f) a nucleotide sequence encoding the CK α -5 polypeptide having the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA clone contained in ATCC Deposit No. 209231; (g) a nucleotide sequence encoding the mature CK α -5 polypeptide having the amino acid sequence encoded by the cDNA clone contained in ATCC Deposit No. 209231; (h) a nucleotide sequence encoding the extracellular domain of the CK α -5 polypeptide having the amino acid sequence encoded by the cDNA clone contained in ATCC Deposit No. 209231; and (i) a nucleotide sequence complementary to any of the nucleotide sequences in (a), (b), (c), (d), (e), (f), (g) or (h) above.

Further embodiments of the invention include isolated nucleic acid molecules that comprise a polynucleotide having a nucleotide sequence at least 90% identical, and more preferably at least 95%, 96%, 97%, 98% or 99% identical, to any of the nucleotide sequences in (a), (b), (c), (d), (e), (f), (g), (h) or (i) above, or a polynucleotide which hybridizes under stringent hybridization conditions to a polynucleotide in (a), (b), (c), (d), (e), (f), (g), (h) or (i), above. This polynucleotide which hybridizes does not hybridize under stringent hybridization conditions to a polynucleotide having a nucleotide sequence consisting of only A residues or of only T residues. An additional nucleic acid embodiment of the invention relates to an isolated nucleic acid molecule comprising a polynucleotide which encodes the amino acid sequence of an epitope-bearing portion of a CK α -5 polypeptide having an amino acid sequence in (a), (b), (c), (d), (e), (f), (g) or (h) above.

The present invention also relates to recombinant vectors, which include the isolated nucleic acid molecules of the present invention, and to host cells containing the recombinant vectors, as well as to methods of making such vectors and host cells and for using them for production of CK α -5 polypeptides or peptides by recombinant techniques.

5 By a polynucleotide having a nucleotide sequence at least, for example, 95% "identical" to a reference nucleotide sequence encoding a CK α -5 polypeptide is intended that the nucleotide sequence of the polynucleotide is identical to the reference sequence except that the polynucleotide sequence may include up to five point mutations per each 100 nucleotides of the reference nucleotide sequence encoding the CK α -5 polypeptide. In other words, to obtain a polynucleotide
10 having a nucleotide sequence at least 95% identical to a reference nucleotide sequence, up to 5% of the nucleotides in the reference sequence may be deleted or substituted with another nucleotide, or a number of nucleotides up to 5% of the total nucleotides in the reference sequence may be inserted into the reference sequence. These mutations of the reference sequence may occur at the 5' or 3' terminal positions of the reference nucleotide sequence or anywhere between those
15 terminal positions, interspersed either individually among nucleotides in the reference sequence or in one or more contiguous groups within the reference sequence.

As a practical matter, whether any particular nucleic acid molecule is at least 90%, 95%, 96%, 97%, 98% or 99% identical to, for instance, the nucleotide sequence shown in Figure 1 or to the nucleotides sequence of the deposited cDNA clone can be determined conventionally using
20 known computer programs such as the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711). Bestfit uses the local homology algorithm of Smith and Waterman, *Advances in Applied Mathematics* 2:482-489 (1981), to find the best segment of homology between two sequences. When using Bestfit or any other sequence alignment program to
25 determine whether a particular sequence is, for instance, 95% identical to a reference sequence according to the present invention, the parameters are set, of course, such that the percentage of identity is calculated over the full length of the reference nucleotide sequence and that gaps in homology of up to 5% of the total number of nucleotides in the reference sequence are allowed.

The present application is directed to nucleic acid molecules at least 90%, 95%, 96%,
30 97%, 98% or 99% identical to the nucleic acid sequence shown in Figure 1 (SEQ ID NO:1) or to the nucleic acid sequence of the deposited cDNA, irrespective of whether they encode a polypeptide having CK α -5 activity. This is because even where a particular nucleic acid molecule does not encode a polypeptide having CK α -5 activity, one of skill in the art would still know how to use the nucleic acid molecule, for instance, as a hybridization probe or a polymerase chain
35 reaction (PCR) primer. Uses of the nucleic acid molecules of the present invention that do not encode a polypeptide having CK α -5 activity include, inter alia, (1) isolating the CK α -5 gene or allelic variants thereof in a cDNA library; (2) *in situ* hybridization (e.g., "FISH") to metaphase

chromosomal spreads to provide precise chromosomal location of the CK α -5 gene, as described in Verma et al., Human Chromosomes: A Manual of Basic Techniques, Pergamon Press, New York (1988); and Northern Blot analysis for detecting CK α -5 mRNA expression in specific tissues.

5 Preferred, however, are nucleic acid molecules having sequences at least 90%, 95%, 96%, 97%, 98% or 99% identical to the nucleic acid sequence shown in Figure 1 (SEQ ID NO:1) or to the nucleic acid sequence of the deposited cDNA which do, in fact, encode a polypeptide having CK α -5 protein activity. By "a polypeptide having CK α -5 activity" is intended polypeptides exhibiting activity similar, but not necessarily identical, to an activity of the
10 extracellular portion of the complete CK α -5 protein. One such activity believed to be possessed by such polypeptides is the ability to modulate colony formation of bone marrow progenitor cells, as does Human Chemokine HCC-1. An in vitro colony forming assay for measuring the extent of inhibition of myeloid progenitor cells is described in Young et al., *The Journal of Immunology* 155:2661-2667 (1995). Briefly, the assay involves collecting human or mouse bone marrow
15 cells and plating the same on agar, adding one or more growth factors and either (1) transfected host cell-supernatant containing CK α -5 protein (or a candidate polypeptide) or (2) nontransfected host cell-supernatant control, and measuring the effect on colony formation by murine and human CFU-granulocyte-macrophages (CFU-GM), by human burst-forming unit-erythroid (BFU-E), or by human CFU granulocyte-erythroid-macrophage-megakaryocyte (CFU-GEMM). Such activity
20 is useful for protecting myeloid progenitor cells during chemotherapy.

CK α -5 protein modulates immune system cell proliferation and differentiation in a dose-dependent manner in the above-described assay. Thus, "a polypeptide having CK α -5 protein activity" includes polypeptides that also exhibit any of the same myeloid progenitor modulation activities in the above-described assays in a dose-dependent manner. Although the degree of
25 dose-dependent activity need not be identical to that of the CK α -5 protein, preferably, "a polypeptide having CK α -5 protein activity" will exhibit substantially similar dose-dependence in a given activity as compared to the CK α -5 protein (i.e., the candidate polypeptide will exhibit greater activity or not more than about 25-fold less and, preferably, not more than about tenfold less activity relative to the reference CK α -5 protein).

30 Like other CXC chemokines, CK α -5 exhibits activity on leukocytes including for example monocytes, lymphocytes and neutrophils. For this reason CK α -5 is active in directing the proliferation, differentiation and migration of these cell types. Such activity is useful for immune enhancement or suppression, myeloprotection, stem cell mobilization, acute and chronic inflammatory control and treatment of leukemia. Assays for measuring such activity are known

in the art. For example, see Peters *et al.*, *Immun. Today* 17:273 (1996); Young *et al.*, *J. Exp. Med.* 182:1111 (1995); Caux *et al.*, *Nature* 390:258 (1992); and Santiago-Schwarz *et al.*, *Adv. Exp. Med. Biol.* 378:7 (1995)." The assays and protocols from each of the forgoing journal articles are incorporated herein by reference.

5 Of course, due to the degeneracy of the genetic code, one of ordinary skill in the art will immediately recognize that a large number of the nucleic acid molecules having a sequence at least 90%, 95%, 96%, 97%, 98%, or 99% identical to the nucleic acid sequence of the deposited cDNA or the nucleic acid sequence shown in Figure 1 (SEQ ID NO:1) will encode a polypeptide "having CK α -5 protein activity." In fact, since degenerate variants of these nucleotide sequences
10 all encode the same polypeptide, this will be clear to the skilled artisan even without performing the above described comparison assay. It will be further recognized in the art that, for such nucleic acid molecules that are not degenerate variants, a reasonable number will also encode a polypeptide having CK α -5 protein activity. This is because the skilled artisan is fully aware of amino acid substitutions that are either less likely or not likely to significantly effect protein
15 function (e.g., replacing one aliphatic amino acid with a second aliphatic amino acid), as further described below.

Vectors and Host Cells

The present invention also relates to vectors which include the isolated DNA molecules of the present invention, host cells which are genetically engineered with the recombinant vectors,
20 and the production of CK α -5 polypeptides or fragments thereof by recombinant techniques. The vector may be, for example, a phage, plasmid, viral or retroviral vector. Retroviral vectors may be replication competent or replication defective. In the latter case, viral propagation generally will occur only in complementing host cells.

The polynucleotides may be joined to a vector containing a selectable marker for
25 propagation in a host. Generally, a plasmid vector is introduced in a precipitate, such as a calcium phosphate precipitate, or in a complex with a charged lipid. If the vector is a virus, it may be packaged in vitro using an appropriate packaging cell line and then transduced into host cells.

The DNA insert should be operatively linked to an appropriate promoter, such as the
30 phage lambda PL promoter, the *E. coli lac*, *trp*, *phoA* and *tac* promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name a few. Other suitable promoters will be known to the skilled artisan. The expression constructs will further contain sites for transcription initiation, termination and, in the transcribed region, a ribosome binding site for translation. The coding portion of the transcripts expressed by the constructs will preferably include a translation
35 initiating codon at the beginning and a termination codon (UAA, UGA or UAG) appropriately positioned at the end of the polypeptide to be translated.

As indicated, the expression vectors will preferably include at least one selectable marker. Such markers include dihydrofolate reductase, G418 or neomycin resistance for eukaryotic cell culture and tetracycline, kanamycin or ampicillin resistance genes for culturing in *E. coli* and other bacteria. Representative examples of appropriate hosts include, but are not limited to, bacterial cells, such as *E. coli*, *Streptomyces* and *Salmonella typhimurium* cells; fungal cells, such as yeast cells; insect cells such as *Drosophila* S2 and *Spodoptera* Sf9 cells; animal cells such as CHO, COS, 293 and Bowes melanoma cells; and plant cells. Appropriate culture mediums and conditions for the above-described host cells are known in the art.

Among vectors preferred for use in bacteria include pQE70, pQE60 and pQE-9, available from QIAGEN, Inc., *supra*; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16a, pNH18A, pNH46A, available from Stratagene; and ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 available from Pharmacia. Among preferred eukaryotic vectors are pWLNEO, pSV2CAT, pOG44, pXT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. Other suitable vectors will be readily apparent to the skilled artisan.

Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-dextran mediated transfection, cationic lipid-mediated transfection, electroporation, transduction, infection or other methods. Such methods are described in many standard laboratory manuals, such as Davis *et al.*, *Basic Methods In Molecular Biology* (1986).

The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals, but also additional heterologous functional regions. For instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and persistence in the host cell, during purification, or during subsequent handling and storage. Also, peptide moieties may be added to the polypeptide to facilitate purification. Such regions may be removed prior to final preparation of the polypeptide. The addition of peptide moieties to polypeptides to engender secretion or excretion, to improve stability and to facilitate purification, among others, are familiar and routine techniques in the art. A preferred fusion protein comprises a heterologous region from immunoglobulin that is useful to stabilize and purify proteins. For example, EP-A-O 464 533 (Canadian counterpart 2045869) discloses fusion proteins comprising various portions of constant region of immunoglobulin molecules together with another human protein or part thereof. In many cases, the Fc part in a fusion protein is thoroughly advantageous for use in therapy and diagnosis and thus results, for example, in improved pharmacokinetic properties (EP-A 0232 262). On the other hand, for some uses it would be desirable to be able to delete the Fc part after the fusion protein has been expressed, detected and purified in the advantageous manner described. This is the case when Fc portion proves to be a hindrance to use in therapy and diagnosis, for example when the fusion protein is to be used as antigen for immunizations. In drug discovery, for example, human proteins, such as hIL-5, have been fused with Fc portions for the purpose of high-throughput screening assays to identify antagonists of hIL-5.

See, D. Bennett *et al.*, *J. Molecular Recognition* 8:52-58 (1995) and K. Johanson *et al.*, *J. Biol. Chem.* 270:9459-9471 (1995).

The CK α -5 protein can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Most preferably, high performance liquid chromatography ("HPLC") is employed for purification. Polypeptides of the present invention include: products purified from natural sources, including bodily fluids, tissues and cells, whether directly isolated or cultured; products of chemical synthetic procedures; and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes. Thus, it is well known in the art that the N-terminal methionine encoded by the translation initiation codon generally is removed with high efficiency from any protein after translation in all eukaryotic cells. While the N-terminal methionine on most proteins also is efficiently removed in most prokaryotes, for some proteins this prokaryotic removal process is inefficient, depending on the nature of the amino acid to which the N-terminal methionine is covalently linked.

Polypeptides and Fragments

The invention further provides an isolated CK α -5 polypeptide having the amino acid sequence encoded by the deposited cDNA, or the amino acid sequence in SEQ ID NO:2, or a peptide or polypeptide comprising a portion of the above polypeptides.

Variant and Mutant Polypeptides

To improve or alter the characteristics of CK α -5 polypeptides, protein engineering may be employed. Recombinant DNA technology known to those skilled in the art can be used to create novel mutant proteins or "muteins including single or multiple amino acid substitutions, deletions, additions or fusion proteins. Such modified polypeptides can show, e.g., enhanced activity or increased stability. In addition, they may be purified in higher yields and show better solubility than the corresponding natural polypeptide, at least under certain purification and storage conditions.

N-Terminal and C-Terminal Deletion Mutants

For instance, for many proteins, including the extracellular domain of a membrane associated protein or the mature form(s) of a secreted protein, it is known in the art that one or

more amino acids may be deleted from the N-terminus or C-terminus without substantial loss of biological function. For instance, Ron et al., *J. Biol. Chem.*, 268:2984-2988 (1993) reported modified KGF proteins that had heparin binding activity even if 3, 8, or 27 amino-terminal amino acid residues were missing. In the present case, since the protein of the invention is a member of the chemokine polypeptide family, deletions of N-terminal amino acids up to the first "Cys" required for formation of a disulfide bridge may retain some biological activity such as receptor binding or modulation of target cell activities. Polypeptides having further N-terminal deletions including the cysteine residue at position 38 in Figure 1 (position 11 in SEQ ID NO:2) would not be expected to retain such biological activities because it is known that this residue in a chemokine-related polypeptide is required for forming a disulfide bridge to provide structural stability which is needed for receptor binding and signal transduction.

However, even if deletion of one or more amino acids from the N-terminus of a protein results in modification or loss of one or more biological functions of the protein, other biological activities may still be retained. Thus, the ability of the shortened protein to induce and/or bind to antibodies which recognize the complete or mature form of the protein generally will be retained when less than the majority of the residues of the complete or mature protein are removed from the N-terminus. Whether a particular polypeptide lacking N-terminal residues of a complete protein retains such immunologic activities can readily be determined by routine methods described herein and otherwise known in the art.

Accordingly, the present invention further provides polypeptides having one or more residues deleted from the amino terminus of the amino acid sequence of the CK α -5 protein shown in SEQ ID NO:2, up to the cysteine residue at position number 11, and polynucleotides encoding such polypeptides. In particular, the present invention provides polypeptides comprising the amino acid sequence of residues n-178 of SEQ ID NO:2, where n is an integer in the range of -4 to 11, and Cys-11 is the position of the first residue from the N-terminus of the extracellular domain of the CK α -5 polypeptide (shown in SEQ ID NO:2) believed to be required for receptor binding activity of the CK α -5 protein.

More in particular, the invention provides polypeptides comprising an amino acid sequence selected from the group consisting of: -4 to 178, -3 to 178, -2 to 178, -1 to 178, 1 to 178, 2 to 178, 3 to 178, 4 to 178, 5 to 178, 6 to 178, 7 to 178, 8 to 178, 9 to 178, 10 to 178, and 11 to 178, all of SEQ ID NO:2. Polynucleotides encoding such polypeptides are also provided.

Similarly, many examples of biologically functional C-terminal deletion muteins are known. For instance, Interferon gamma shows up to ten times higher activities by deleting 8-10 amino acid residues from the carboxy terminus of the protein (Döbeli et al., *J. Biotechnology* 7:199-216 (1988)). In the present case, since the protein of the invention is a member of the chemokine polypeptide family, deletions of C-terminal amino acids up to the cysteine at position 55 of SEQ ID NO:2 may retain some biological activity such as receptor binding or modulation of target cell activities. Polypeptides having further C-terminal deletions including the cysteine

residue at position 82 of Figure 1 (55 of SEQ ID NO:2) would not be expected to retain such biological activities because it is known that this residue in a chemokine-related polypeptide is required for forming a disulfide bridge to provide structural stability which is needed for receptor binding and signal transduction.

5 However, even if deletion of one or more amino acids from the C-terminus of a protein results in modification or loss of one or more biological functions of the protein, other biological activities may still be retained. Thus, the ability of the shortened protein to induce and/or bind to antibodies which recognize the complete or mature form of the protein generally will be retained when less than the majority of the residues of the complete or mature protein are removed from
10 the C-terminus. Whether a particular polypeptide lacking C-terminal residues of a complete protein retains such immunologic activities can readily be determined by routine methods described herein and otherwise known in the art.

Accordingly, the present invention further provides polypeptides having one or more residues from the carboxy terminus of the amino acid sequence of the CK α -5 shown in SEQ ID
15 NO:2, up to the cysteine residue at position 55 of SEQ ID NO:2, and polynucleotides encoding such polypeptides. In particular, the present invention provides polypeptides having the amino acid sequence of residues (-4)-m of the amino acid sequence in SEQ ID NO:2, where m is any integer in the range of 55 to 178, and residue cysteine-55 is the position of the first residue from the C-terminus of the complete CK α -5 polypeptide (shown in SEQ ID NO:2) believed to be
20 required for receptor binding and target cell modulation activities of the CK α -5 protein.

More in particular, the invention provides polypeptides comprising an amino acid sequence selected from the group consisting of: -4 to 55, -4 to 56, -4 to 57, -4 to 58, -4 to 59, -4 to 60, -4 to 61, -4 to 62, -4 to 63, -4 to 64, -4 to 65, -4 to 66, -4 to 67, -4 to 68, -4 to 69, -4 to 70, -4 to 71, -4 to 72, -4 to 73, -4 to 74, -4 to 75, -4 to 76, -4 to 77, -4 to 78, -4 to 79, -4 to 79,
25 -4 to 80, -4 to 81, -4 to 82, -4 to 83, -4 to 84, -4 to 85, -4 to 86, -4 to 87, -4 to 88, -4 to 89, -4 to 90, -4 to 91, -4 to 92, -4 to 93, -4 to 94, -4 to 95, -4 to 96, -4 to 95, -4 to 96, -4 to 97, -4 to 98, -4 to 99, -4 to 100, -4 to 101, -4 to 102, -4 to 103, -4 to 104, -4 to 105, -4 to 106, -4 to 107, -4 to 108, -4 to 109, -4 to 110, -4 to 111, -4 to 112, -4 to 113, -4 to 114, -4 to 115, -4 to 116, -4 to 117, -4 to 118, -4 to 119, -4 to 120, -4 to 121, -4 to 122, -4 to 123, -4 to 124, -4 to 125, -4 to
30 126, -4 to 127, -4 to 128, -4 to 129, -4 to 130, -4 to 131, -4 to 132, -4 to 133, -4 to 134, -4 to 135, -4 to 136, -4 to 137, -4 to 138, -4 to 139, -4 to 140, -4 to 141, -4 to 142, -4 to 143, -4 to 144, -4 to 145, -4 to 146, -4 to 147, -4 to 148, -4 to 149, -4 to 150, -4 to 151, -4 to 152, -4 to 153, -4 to 154, -4 to 155, -4 to 156, -4 to 157, -4 to 158, -4 to 159, -4 to 160, -4 to 161, -4 to 162, -4 to 163, -4 to 164, -4 to 165, -4 to 166, -4 to 167, -4 to 168, -4 to 169, -4 to 170, -4 to
35 171, -4 to 172, -4 to 173, -4 to 174, -4 to 175, -4 to 176, -4 to 177 and -4 to 178, all of SEQ ID NO:2. Polynucleotides encoding these polypeptides also are provided.

The invention also provides polypeptides having one or more amino acids deleted from both the amino and the carboxyl termini of the soluble form (extracellular domain), which may be

described generally as having residues n-m of SEQ ID NO:2, where n and m are integers as described above. Particularly preferred are polypeptides comprising an amino acid sequence selected from the group consisting of: 11 to 55, 11 to 56, 10 to 57, 10 to 58, 10 to 59, 10 to 60, 10 to 61, 10 to 62, 9 to 63, 9 to 64, 9 to 65, 9 to 66, 9 to 67, 9 to 68, 9 to 69, 9 to 70, 9 to 71, 9 to 72, 9 to 73, 8 to 74, 8 to 75, 8 to 76, 8 to 77, 8 to 78, 8 to 79, 8 to 80, 8 to 81, 8 to 82, 8 to 83, 8 to 84, 7 to 85, 7 to 86, 7 to 87, 7 to 88, 7 to 89, 7 to 90, 7 to 91, 7 to 92, 7 to 93, 7 to 94, 7 to 95, 7 to 96, 7 to 95, 7 to 96, 7 to 97, 7 to 98, 7 to 99, 7 to 100, 6 to 101, 6 to 102, 6 to 103, 6 to 104, 6 to 105, 6 to 106, 6 to 107, 6 to 108, 6 to 109, 6 to 110, 6 to 111, 6 to 112, 6 to 113, 6 to 114, 6 to 115, 6 to 116, 6 to 117, 6 to 118, 6 to 119, 6 to 120, 6 to 121, 6 to 122, 6 to 123, 6 to 124, 6 to 125, 6 to 126, 6 to 127, 6 to 128, 6 to 129, 6 to 130, 6 to 131, 6 to 132, 6 to 133, 6 to 134, 6 to 135, 6 to 136, 6 to 137, 6 to 138, 6 to 139, 6 to 140, 6 to 141, 6 to 142, 6 to 143, 6 to 144, 6 to 145, 6 to 146, 6 to 147, 6 to 148, 6 to 149, 6 to 150, 6 to 151, 6 to 152, 6 to 153, 6 to 154, 6 to 155, 6 to 156, 6 to 157, 5 to 158, 5 to 159, 5 to 160, 5 to 161, 5 to 162, 5 to 163, 5 to 164, 5 to 165, 5 to 166, 5 to 167, 5 to 168, 5 to 169, 4 to 170, 4 to 171, 4 to 172, 4 to 173, 4 to 174, 4 to 175, 4 to 176, 4 to 177 and 4 to 178, all of SEQ ID NO:2.

Polynucleotides encoding these polypeptides also are provided.

Also included are a nucleotide sequence encoding a polypeptide consisting of a portion of the complete CK α -5 amino acid sequence encoded by the cDNA clone contained in ATCC Deposit No. 209231, where this portion excludes from 23 to about 37 amino acids from the amino terminus of the extracellular domain of the CK α -5 polypeptide encoded by the cDNA clone contained in ATCC Deposit No. 209231, or from 49 to about 172 amino acids from the carboxy terminus, or any combination of the above amino terminal and carboxy terminal deletions, of the extracellular domain encoded by the cDNA clone contained in ATCC Deposit No. 209231. Polynucleotides encoding all of the above deletion mutant polypeptide forms also are provided.

Other Mutants

In addition to terminal deletion forms of the protein discussed above, it also will be recognized by one of ordinary skill in the art that some amino acid sequences of the CK α -5 polypeptide can be varied without significant effect of the structure or function of the protein. If such differences in sequence are contemplated, it should be remembered that there will be critical areas on the protein which determine activity.

Thus, the invention further includes variations of the CK α -5 polypeptide which show substantial CK α -5 polypeptide activity or which include regions of CK α -5 protein such as the protein portions discussed below. Such mutants include deletions, insertions, inversions, repeats, and type substitutions selected according to general rules known in the art so as have little effect on activity. For example, guidance concerning how to make phenotypically silent amino acid substitutions is provided in Bowie, J. U. et al., "Deciphering the Message in Protein

Sequences: Tolerance to Amino Acid Substitutions," *Science* 247:1306-1310 (1990), wherein the authors indicate that there are two main approaches for studying the tolerance of an amino acid sequence to change. The first method relies on the process of evolution, in which mutations are either accepted or rejected by natural selection. The second approach uses genetic engineering to introduce amino acid changes at specific positions of a cloned gene and selections or screens to identify sequences that maintain functionality.

As the authors state, these studies have revealed that proteins are surprisingly tolerant of amino acid substitutions. The authors further indicate which amino acid changes are likely to be permissive at a certain position of the protein. For example, most buried amino acid residues require nonpolar side chains, whereas few features of surface side chains are generally conserved. Other such phenotypically silent substitutions are described in Bowie, J. U. *et al.*, *supra*, and the references cited therein. Typically seen as conservative substitutions are those shown in Table 1.

Thus, the fragment, derivative or analog of the polypeptide of SEQ ID NO:2, or that encoded by the deposited cDNA, may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the above form of the polypeptide, such as an IgG Fc fusion region peptide or leader or secretory sequence or a sequence which is employed for purification of the above form of the polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art from the teachings herein

Thus, the CK α -5 of the present invention may include one or more amino acid substitutions, deletions or additions, either from natural mutations or human manipulation. As indicated, changes are preferably of a minor nature, such as conservative amino acid substitutions that do not significantly affect the folding or activity of the protein (see Table 1).

TABLE 1. Conservative Amino Acid Substitutions.

Aromatic	Phenylalanine Tryptophan Tyrosine
Hydrophobic	Leucine Isoleucine Valine
Polar	Glutamine Asparagine
Basic	Arginine Lysine Histidine
Acidic	Aspartic Acid Glutamic Acid
Small	Alanine Serine Threonine Methionine Glycine

Amino acids in the CK α -5 protein of the present invention that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science* 244:1081-1085 (1989)). The latter procedure introduces single alanine mutations at every residue in the molecule. The resulting mutant molecules are then tested for biological activity such as receptor binding or *in vitro* or *in vitro* proliferative activity.

Of special interest are substitutions of charged amino acids with other charged or neutral amino acids which may produce proteins with highly desirable improved characteristics, such as less aggregation. Aggregation may not only reduce activity but also be problematic when preparing pharmaceutical formulations, because aggregates can be immunogenic (Pinckard *et al.*, *Clin. Exp. Immunol.* 2:331-340 (1967); Robbins *et al.*, *Diabetes* 36: 838-845 (1987); Cleland *et al.*, *Crit. Rev. Therapeutic Drug Carrier Systems* 10:307-377 (1993).

Replacement of amino acids can also change the selectivity of the binding of a ligand to cell surface receptors. For example, Ostade *et al.*, *Nature* 361:266-268 (1993) describes certain

mutations resulting in selective binding of TNF- α to only one of the two known types of TNF receptors. Sites that are critical for ligand-receptor binding can also be determined by structural analysis such as crystallization, nuclear magnetic resonance or photoaffinity labeling (Smith *et al.*, *J. Mol. Biol.* 224:899-904 (1992) and de Vos *et al. Science* 255:306-312 (1992)).

5 As described above CK α -5 lacks an ELR motif. By making a specific mutation in CK α -5 to include an ELR motif in the position where such a motif is typically found in related chemokines, CK α -5 will act as an antagonist, thus possessing angiogenic activity. Accordingly, polypeptides of the present invention include CK α -5-ELR mutants. Such CK α -5-ELR mutants are comprised of the full-length or preferably the mature CK α -5 protein. Further, since CK α -5
10 is a member of the chemokine-related protein family, to modulate rather than completely eliminate biological activities of CK α -5 preferably mutations are made in sequences encoding amino acids in the CK α -5 extracellular domain, i.e., in positions 1 to 178 of SEQ ID NO:2, more preferably in residues within this region which are not conserved in all members of the chemokine family. As is known in the art, the four spatially conserved cyteines present in all chemokines, positions
15 11, 13, 41 and 55 in CK α -5, are required for the formation of two disulfide bridges. Thus, it is preferable not to alter any of the four cysteine residues located at positions 11, 13, 41 and 55 in SEQ ID NO:2. Also forming part of the present invention are isolated polynucleotides comprising nucleic acid sequences which encode the above CK α -5 mutants.

 The polypeptides of the present invention are preferably provided in an isolated form, and
20 preferably are substantially purified. A recombinantly produced version of the CK α -5 polypeptide can be substantially purified by the one-step method described in Smith and Johnson, *Gene* 67:31-40 (1988). Polypeptides of the invention also can be purified from natural or recombinant sources using anti-CK α -5 antibodies of the invention in methods which are well known in the art of protein purification.

The invention further provides an isolated CK α -5 polypeptide comprising an amino acid sequence selected from the group consisting of: (a) the amino acid sequence of the full-length CK α -5 polypeptide having the complete amino acid sequence shown in SEQ ID NO:2 or the complete amino acid sequence encoded by the cDNA clone contained in the ATCC Deposit No. 209231; (b) the amino acid sequence of the full-length CK α -5 polypeptide having the complete amino acid sequence shown in SEQ ID NO:2 excepting the N-terminal methionine (i.e., positions -26 to 227 of SEQ ID NO:2) or the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA clone contained in the ATCC Deposit No. 209231; (c) the amino acid sequence of the mature CK α -5 polypeptide having the amino acid sequence shown in SEQ ID NO:2 as residues 1-227 or the amino acid sequence of the mature CK α -5 encoded by the cDNA clone contained in the ATCC Deposit No. 209231; and (d) the amino acid sequence of the extracellular domain of the CK α -5 polypeptide having the amino acid sequence shown in SEQ ID NO:2 as residues 1-178 or the amino acid sequence of the extracellular domain of the CK α -5 polypeptide encoded by the cDNA clone contained in the ATCC Deposit No. 209231.

Further polypeptides of the present invention include polypeptides which have at least 90% similarity, more preferably at least 95% similarity, and still more preferably at least 96%, 97%, 98% or 99% similarity to those described above. The polypeptides of the invention also comprise those which are at least 80% identical, more preferably at least 90% or 95% identical, still more preferably at least 96%, 97%, 98% or 99% identical to the polypeptide encoded by the deposited cDNA or to the polypeptide of SEQ ID NO:2, and also include portions of such polypeptides with at least 30 amino acids and more preferably at least 50 amino acids. Polynucleotides encoding such variant polypeptides are also provided.

By "% similarity" for two polypeptides is intended a similarity score produced by comparing the amino acid sequences of the two polypeptides using the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711) and the default settings for determining similarity. Bestfit uses the local homology algorithm of Smith and Waterman (Advances in Applied Mathematics 2:482-489, 1981) to find the best segment of similarity between two sequences.

By a polypeptide having an amino acid sequence at least, for example, 95% "identical" to a reference amino acid sequence of a CK α -5 polypeptide is intended that the amino acid sequence of the polypeptide is identical to the reference sequence except that the polypeptide sequence may include up to five amino acid alterations per each 100 amino acids of the reference amino acid of the CK α -5 polypeptide. In other words, to obtain a polypeptide having an amino acid sequence at least 95% identical to a reference amino acid sequence, up to 5% of the amino acid residues in

the reference sequence may be deleted or substituted with another amino acid, or a number of amino acids up to 5% of the total amino acid residues in the reference sequence may be inserted into the reference sequence. These alterations of the reference sequence may occur at the amino or carboxy terminal positions of the reference amino acid sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in one or more contiguous groups within the reference sequence.

As a practical matter, whether any particular polypeptide is at least 90%, 95%, 96%, 97%, 98% or 99% identical to, for instance, the amino acid sequence shown in SEQ ID NO:2 or to the amino acid sequence encoded by deposited cDNA clone can be determined conventionally using known computer programs such the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711). When using Bestfit or any other sequence alignment program to determine whether a particular sequence is, for instance, 95% identical to a reference sequence according to the present invention, the parameters are set, of course, such that the percentage of identity is calculated over the full length of the reference amino acid sequence and that gaps in homology of up to 5% of the total number of amino acid residues in the reference sequence are allowed.

The polypeptide of the present invention could be used as a molecular weight marker on SDS-PAGE gels or on molecular sieve gel filtration columns using methods well known to those of skill in the art.

As described in detail below, the polypeptides of the present invention can also be used to raise polyclonal and monoclonal antibodies, which are useful in assays for detecting CK α -5 protein expression as described below or as agonists and antagonists capable of enhancing or inhibiting CK α -5 protein function. Further, such polypeptides can be used in the yeast two-hybrid system to "capture" CK α -5 protein binding proteins which are also candidate agonists and antagonists according to the present invention. The yeast two hybrid system is described in Fields and Song, Nature 340:245-246 (1989).

Epitope-Bearing Portions

In another aspect, the invention provides a peptide or polypeptide comprising an epitope-bearing portion of a polypeptide of the invention. The epitope of this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide of the invention. An "immunogenic epitope" is defined as a part of a protein that elicits an antibody response when the whole protein is the immunogen. On the other hand, a region of a protein molecule to which an antibody can bind is defined as an "antigenic epitope." The number of immunogenic epitopes of a protein generally is less than the number of antigenic epitopes. See, for instance, Geysen *et al.*, *Proc. Natl. Acad. Sci. USA* 81:3998-4002 (1983).

As to the selection of peptides or polypeptides bearing an antigenic epitope (i.e., that contain a region of a protein molecule to which an antibody can bind), it is well known in that art that relatively short synthetic peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See, for instance, 5 Sutcliffe, J. G., Shinnick, T. M., Green, N. and Learner, R. A. (1983) "Antibodies that react with predetermined sites on proteins," *Science*, 219:660-666. Peptides capable of eliciting protein-reactive sera are frequently represented in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined neither to immunodominant regions of intact proteins (i.e., immunogenic epitopes) nor to the amino or carboxyl terminals. 10 Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. See, for instance, Wilson *et al.*, *Cell* 37:767-778 (1984) at 777.

Antigenic epitope-bearing peptides and polypeptides of the invention preferably contain a sequence of at least seven, more preferably at least nine and most preferably between 15 about 15 to about 30 amino acids contained within the amino acid sequence of a polypeptide of the invention. Non-limiting examples of antigenic polypeptides or peptides that can be used to generate CK α -5-specific antibodies include: a polypeptide comprising amino acid residues from about Tyr-12 to about Ser-24 in SEQ ID NO:2; a polypeptide comprising amino acid residues from about Cys-55 to about Val-63 in SEQ ID NO:2; a polypeptide comprising amino acid 20 residues from about Thr-91 to about Pro-110 in SEQ ID NO:2; and a polypeptide comprising amino acid residues from about Ser-120 to about Ile-145. These polypeptide fragments have been determined to bear antigenic epitopes of the CK α -5 protein by the analysis of the Jameson-Wolf antigenic index, as shown in Figure 3, above.

The epitope-bearing peptides and polypeptides of the invention may be produced by any 25 conventional means. See, e.g., Houghten, R. A. (1985) "General method for the rapid solid-phase synthesis of large numbers of peptides: specificity of antigen-antibody interaction at the level of individual amino acids." *Proc. Natl. Acad. Sci. USA* 82:5131-5135; this "Simultaneous Multiple Peptide Synthesis (SMPS)" process is further described in U.S. Patent No. 4,631,211 to Houghten *et al.* (1986).

30 Epitope-bearing peptides and polypeptides of the invention are used to induce antibodies according to methods well known in the art. See, for instance, Sutcliffe *et al.*, *supra*; Wilson *et al.*, *supra*; Chow, M. *et al.*, *Proc. Natl. Acad. Sci. USA* 82:910-914; and Bittle, F. J. *et al.*, *J. Gen. Virol.* 66:2547-2354 (1985). Immunogenic epitope-bearing peptides of the invention, i.e., those parts of a protein that elicit an antibody response when the whole protein is the immunogen, 35 are identified according to methods known in the art. See, for instance, Geysen *et al.*, *supra*. Further still, U.S. Patent No. 5,194,392 to Geysen (1990) describes a general method of detecting or determining the sequence of monomers (amino acids or other compounds) which is a topological equivalent of the epitope (i.e., a "mimotope") which is complementary to a particular paratope (antigen binding site) of an antibody of interest. More generally, U.S. Patent No.

4,433,092 to Geysen (1989) describes a method of detecting or determining a sequence of monomers which is a topographical equivalent of a ligand which is complementary to the ligand binding site of a particular receptor of interest. Similarly, U.S. Patent No. 5,480,971 to Houghten, R. A. et al. (1996) on Peralkylated Oligopeptide Mixtures discloses linear
5 C1-C7-alkyl peralkylated oligopeptides and sets and libraries of such peptides, as well as methods for using such oligopeptide sets and libraries for determining the sequence of a peralkylated oligopeptide that preferentially binds to an acceptor molecule of interest. Thus, non-peptide analogs of the epitope-bearing peptides of the invention also can be made routinely by these methods.

10 ***Fusion Proteins***

As one of skill in the art will appreciate, CK α -5 polypeptides of the present invention and the epitope-bearing fragments thereof described above can be combined with parts of the constant domain of immunoglobulins (IgG), resulting in chimeric polypeptides. These fusion proteins facilitate purification and show an increased half-life in vivo. This has been shown, e.g., for
15 chimeric proteins consisting of the first two domains of the human CD4-polypeptide and various domains of the constant regions of the heavy or light chains of mammalian immunoglobulins (EP A 394,827; Traunecker et al., *Nature* 331:84-86 (1988)). Fusion proteins that have a disulfide-linked dimeric structure due to the IgG part can also be more efficient in binding and neutralizing other molecules than the monomeric CK α -5 protein or protein fragment alone
20 (Fountoulakis et al., *J. Biochem.* 270:3958-3964 (1995)).

Antibodies

CK α -5-protein specific antibodies for use in the present invention can be raised against the intact CK α -5 protein or an antigenic polypeptide fragment thereof, which may be presented together with a carrier protein, such as an albumin, to an animal system (such as rabbit or mouse)
25 or, if it is long enough (at least about 25 amino acids), without a carrier.

As used herein, the term "antibody" (Ab) or "monoclonal antibody" (Mab) is meant to include intact molecules as well as antibody fragments (such as, for example, Fab and F(ab')₂ fragments) which are capable of specifically binding to CK α -5 protein. Fab and F(ab')₂ fragments lack the Fc fragment of intact antibody, clear more rapidly from the circulation, and
30 may have less non-specific tissue binding of an intact antibody (Wahl et al., *J. Nucl. Med.* 24:316-325 (1983)). Thus, these fragments are preferred.

The antibodies of the present invention may be prepared by any of a variety of methods. For example, cells expressing the CK α -5 protein or an antigenic fragment thereof can be administered to an animal in order to induce the production of sera containing polyclonal
35 antibodies. In a preferred method, a preparation of CK α -5 protein is prepared and purified to

render it substantially free of natural contaminants. Such a preparation is then introduced into an animal in order to produce polyclonal antisera of greater specific activity.

In the most preferred method, the antibodies of the present invention are monoclonal antibodies (or CK α -5 protein binding fragments thereof). Such monoclonal antibodies can be prepared using hybridoma technology (Köhler *et al.*, *Nature* 256:495 (1975); Köhler *et al.*, *Eur. J. Immunol.* 6:511 (1976); Köhler *et al.*, *Eur. J. Immunol.* 6:292 (1976); Hammerling *et al.*, in: *Monoclonal Antibodies and T-Cell Hybridomas*, Elsevier, N.Y., (1981) pp. 563-681). In general, such procedures involve immunizing an animal (preferably a mouse) with a CK α -5 protein antigen or, more preferably, with a CK α -5 protein-expressing cell. Suitable cells can be recognized by their capacity to bind anti-CK α -5 protein antibody. Such cells may be cultured in any suitable tissue culture medium; however, it is preferable to culture cells in Earle's modified Eagle's medium supplemented with 10% fetal bovine serum (inactivated at about 56° C), and supplemented with about 10 g/l of nonessential amino acids, about 1,000 U/ml of penicillin, and about 100 μ g/ml of streptomycin. The splenocytes of such mice are extracted and fused with a suitable myeloma cell line. Any suitable myeloma cell line may be employed in accordance with the present invention; however, it is preferable to employ the parent myeloma cell line (SP2O), available from the American Type Culture Collection, Rockville, Maryland. After fusion, the resulting hybridoma cells are selectively maintained in HAT medium, and then cloned by limiting dilution as described by Wands *et al.* (*Gastroenterology* 80:225-232 (1981)). The hybridoma cells obtained through such a selection are then assayed to identify clones which secrete antibodies capable of binding the CK α -5 protein antigen.

Alternatively, additional antibodies capable of binding to the CK α -5 protein antigen may be produced in a two-step procedure through the use of anti-idiotypic antibodies. Such a method makes use of the fact that antibodies are themselves antigens, and that, therefore, it is possible to obtain an antibody which binds to a second antibody. In accordance with this method, CK α -5-protein specific antibodies are used to immunize an animal, preferably a mouse. The splenocytes of such an animal are then used to produce hybridoma cells, and the hybridoma cells are screened to identify clones which produce an antibody whose ability to bind to the CK α -5 protein-specific antibody can be blocked by the CK α -5 protein antigen. Such antibodies comprise anti-idiotypic antibodies to the CK α -5 protein-specific antibody and can be used to immunize an animal to induce formation of further CK α -5 protein-specific antibodies.

It will be appreciated that Fab and F(ab')₂ and other fragments of the antibodies of the present invention may be used according to the methods disclosed herein. Such fragments are typically produced by proteolytic cleavage, using enzymes such as papain (to produce Fab

fragments) or pepsin (to produce F(ab')₂ fragments). Alternatively, CK α -5 protein-binding fragments can be produced through the application of recombinant DNA technology or through synthetic chemistry.

For *in vivo* use of anti-CK α -5 in humans, it may be preferable to use "humanized" chimeric monoclonal antibodies. Such antibodies can be produced using genetic constructs derived from hybridoma cells producing the monoclonal antibodies described above. Methods for producing chimeric antibodies are known in the art. See, for review, Morrison, *Science* 229:1202 (1985); Oi et al., *BioTechniques* 4:214 (1986); Cabilly et al., U.S. Patent No. 4,816,567; Taniguchi et al., EP 171496; Morrison et al., EP 173494; Neuberger et al., WO 8601533; Robinson et al., WO 8702671; Boulianne et al., *Nature* 312:643 (1984); Neuberger et al., *Nature* 314:268 (1985).

Immune System-Related Disorders

Diagnosis

The present inventors have discovered that CK α -5 is expressed in neutrophils and monocytes. For a number of immune system-related disorders, substantially altered (increased or decreased) levels of CK α -5 gene expression can be detected in immune system tissue or other cells or bodily fluids (e.g., sera, plasma, urine, synovial fluid or spinal fluid) taken from an individual having such a disorder, relative to a "standard" CK α -5 gene expression level, that is, the CK α -5 expression level in immune system tissues or bodily fluids from an individual not having the immune system disorder. Thus, the invention provides a diagnostic method useful during diagnosis of an immune system disorder, which involves measuring the expression level of the gene encoding the CK α -5 protein in immune system tissue or other cells or body fluid from an individual and comparing the measured gene expression level with a standard CK α -5 gene expression level, whereby an increase or decrease in the gene expression level compared to the standard is indicative of an immune system disorder.

In particular, it is believed that certain tissues in mammals with cancer, particularly osteosarcoma, express significantly altered levels of the CK α -5 protein and mRNA encoding the CK α -5 protein when compared to a corresponding "standard" level. Further, it is believed that altered levels of the CK α -5 protein can be detected in certain body fluids (e.g., sera, plasma, urine, and spinal fluid) from mammals with such a cancer when compared to sera from mammals of the same species not having the cancer.

Thus, the invention provides a diagnostic method useful during diagnosis of an immune system disorder, including cancers of the skeletal and immune systems, which involves

measuring the expression level of the gene encoding the CK α -5 protein in immune or skeletal system tissue or other cells or body fluid from an individual and comparing the measured gene expression level with a standard CK α -5 gene expression level, whereby an increase or decrease in the gene expression level compared to the standard is indicative of an immune system disorder.

Where a diagnosis of a disorder in the immune system, including diagnosis of a tumor, has already been made according to conventional methods, the present invention is useful as a prognostic indicator, whereby patients exhibiting enhanced or depressed CK α -5 gene expression will experience a worse clinical outcome relative to patients expressing the gene at a level nearer the standard level.

By "assaying the expression level of the gene encoding the CK α -5 protein" is intended qualitatively or quantitatively measuring or estimating the level of the CK α -5 protein or the level of the mRNA encoding the CK α -5 protein in a first biological sample either directly (e.g., by determining or estimating absolute protein level or mRNA level) or relatively (e.g., by comparing to the CK α -5 protein level or mRNA level in a second biological sample). Preferably, the CK α -5 protein level or mRNA level in the first biological sample is measured or estimated and compared to a standard CK α -5 protein level or mRNA level, the standard being taken from a second biological sample obtained from an individual not having the disorder or being determined by averaging levels from a population of individuals not having a disorder of the immune system. As will be appreciated in the art, once a standard CK α -5 protein level or mRNA level is known, it can be used repeatedly as a standard for comparison.

By "biological sample" is intended any biological sample obtained from an individual, body fluid, cell line, tissue culture, or other source which contains CK α -5 protein or mRNA. As indicated, biological samples include body fluids (such as sera, plasma, urine, synovial fluid and spinal fluid) which contain free CK α -5 protein, immune and skeletal system tissue, and other tissue sources found to express complete mature CK α -5 or a CK α -5 receptor. Methods for obtaining tissue biopsies and body fluids from mammals are well known in the art. Where the biological sample is to include mRNA, a tissue biopsy is the preferred source.

The present invention is useful for diagnosis or treatment of various immune system-related disorders in mammals, preferably humans. Such disorders include tumors, cancers, sarcomas such as osteosarcoma, interstitial lung disease (such as Langerhans cell granulomatosis) and any dysregulation of immune cell function including, but not limited to, autoimmunity, arthritis, leukemias, lymphomas, immunosuppression, immunity, humoral immunity, inflammatory bowel disease, myelo suppression, and the like.

Total cellular RNA can be isolated from a biological sample using any suitable technique such as the single-step guanidinium-thiocyanate-phenol-chloroform method described in Chomczynski and Sacchi, *Anal. Biochem.* 162:156-159 (1987). Levels of mRNA encoding the CK α -5 protein are then assayed using any appropriate method. These include Northern blot
5 analysis, S1 nuclease mapping, the polymerase chain reaction (PCR), reverse transcription in combination with the polymerase chain reaction (RT-PCR), and reverse transcription in combination with the ligase chain reaction (RT-LCR).

Assaying CK α -5 protein levels in a biological sample can occur using antibody-based techniques. For example, CK α -5 protein expression in tissues can be studied with classical
10 immunohistological methods (Jalkanen, M., *et al.*, *J. Cell. Biol.* 101:976-985 (1985); Jalkanen, M., *et al.*, *J. Cell. Biol.* 105:3087-3096 (1987)). Other antibody-based methods useful for detecting CK α -5 protein gene expression include immunoassays, such as the enzyme linked immunosorbent assay (ELISA) and the radioimmunoassay (RIA). Suitable antibody assay labels are known in the art and include enzyme labels, such as, glucose oxidase, and radioisotopes,
15 such as iodine (^{125}I , ^{121}I), carbon (^{14}C), sulfur (^{35}S), tritium (^3H), indium (^{112}In), and technetium ($^{99\text{m}}\text{Tc}$), and fluorescent labels, such as fluorescein and rhodamine, and biotin.

In addition to assaying CK α -5 protein levels in a biological sample obtained from an individual, CK α -5 protein can also be detected *in vivo* by imaging. Antibody labels or markers for *in vivo* imaging of CK α -5 protein include those detectable by X-radiography, NMR or ESR.
20 For X-radiography, suitable labels include radioisotopes such as barium or cesium, which emit detectable radiation but are not overtly harmful to the subject. Suitable markers for NMR and ESR include those with a detectable characteristic spin, such as deuterium, which may be incorporated into the antibody by labeling of nutrients for the relevant hybridoma.

A CK α -5 protein-specific antibody or antibody fragment which has been labeled with an
25 appropriate detectable imaging moiety, such as a radioisotope (for example, ^{131}I , ^{112}In , $^{99\text{m}}\text{Tc}$), a radio-opaque substance, or a material detectable by nuclear magnetic resonance, is introduced (for example, parenterally, subcutaneously or intraperitoneally) into the mammal to be examined for immune system disorder. It will be understood in the art that the size of the subject and the imaging system used will determine the quantity of imaging moiety needed to produce diagnostic
30 images. In the case of a radioisotope moiety, for a human subject, the quantity of radioactivity injected will normally range from about 5 to 20 millicuries of $^{99\text{m}}\text{Tc}$. The labeled antibody or antibody fragment will then preferentially accumulate at the location of cells which contain CK α -5 protein. *In vivo* tumor imaging is described in S.W. Burchiel et al., "Immunopharmacokinetics of Radiolabeled Antibodies and Their Fragments" (Chapter 13 in *Tumor Imaging: The*

Radiochemical Detection of Cancer, S.W. Burchiel and B. A. Rhodes, eds., Masson Publishing Inc. (1982)).

Treatment

5 As noted above, CK α -5 polynucleotides and polypeptides are useful for diagnosis of conditions involving abnormally high or low expression of CK α -5 activities. Given the cells and tissues where CK α -5 is expressed as well as the activities modulated by CK α -5, it is readily apparent that a substantially altered (increased or decreased) level of expression of CK α -5 in an individual compared to the standard or "normal" level produces pathological conditions related to
10 the bodily system(s) in which CK α -5 is expressed and/or is active.

 It will also be appreciated by one of ordinary skill that, since the CK α -5 protein of the invention is a member of the chemokine family the mature secreted form of the protein may be released in soluble form from the cells which express the CK α -5 by proteolytic cleavage. Therefore, when CK α -5 is added from an exogenous source to cells, tissues or the body of an
15 individual, the protein will exert its physiological activities on its target cells of that individual. Also, cells expressing this transmembrane protein may be added to cells, tissues or the body of an individual and these added cells will bind to cells expressing receptor for CK α -5, whereby the cells expressing CK α -5 can cause actions, e.g., cell stimulation, on the receptor-bearing target cells.

20 Therefore, it will be appreciated that conditions caused by a decrease in the standard or normal level of CK α -5 activity in an individual, particularly disorders of the immune system, can be treated by administration of CK α -5 polypeptide in the form of the soluble extracellular domain or cells expressing the complete protein. Thus, the invention also provides a method of treatment of an individual in need of an increased level of CK α -5 activity comprising administering to such
25 an individual a pharmaceutical composition comprising an amount of an isolated CK α -5 polypeptide of the invention, effective to increase the CK α -5 activity level in such an individual.

 Since CK α -5 lacks an ELR motif, it also should be an angiostatic factor rather than an angiogenic factor. In addition, since CK α -5 inhibits endothelial cell function, it will have a wide range of anti-inflammatory activities. CK α -5 may be employed as an anti-neovascularizing agent
30 to treat solid tumors by stimulating the invasion and activation of host defense cells, e.g., cytotoxic T cells and macrophages and by inhibiting the angiogenesis of tumors. Those of skill in

the art will recognize other non-cancer indications where blood vessel proliferation is not wanted. They may also be employed to enhance host defenses against resistant chronic and acute infections, for example, myobacterial infections via the attraction and activation of microbicidal leukocytes. CK α -5 may also be employed to inhibit T-cell proliferation by the inhibition of IL-2 biosynthesis for the treatment of T-cell mediated auto-immune diseases and lymphocytic leukemias. CK α -5 may also be employed to stimulate wound healing, both via the recruitment of debris clearing and connective tissue promoting inflammatory cells and also via its control of excessive TGF -mediated fibrosis. In this same manner, CK α -5 may also be employed to treat other fibrotic disorders, including liver cirrhosis, osteoarthritis and pulmonary fibrosis. CK α -5 also increases the presence of eosinophils which have the distinctive function of killing the larvae of parasites that invade tissues, as in schistosomiasis, trichinosis and ascariasis. It may also be employed to regulate hematopoiesis, by regulating the activation and differentiation of various hematopoietic progenitor cells, for example, to release mature leukocytes from the bone marrow following chemotherapy, i.e., in stem cell mobilization. CK α -5 may also be employed to treat sepsis. Also, by making a specific mutation to include an ELR motif, CK α -5-ELR will have angiogenic activities which are useful for treating all of the disease states where angiogenesis would be beneficial, i.e., to promote wound healing, re-vascularization of damaged limbs from injury or disease, and others known to those of skill in the art.

20 ***Formulations***

The CK α -5 polypeptide composition will be formulated and dosed in a fashion consistent with good medical practice, taking into account the clinical condition of the individual patient (especially the side effects of treatment with CK α -5 polypeptide alone), the site of delivery of the CK α -5 polypeptide composition, the method of administration, the scheduling of administration, and other factors known to practitioners. The "effective amount" of CK α -5 polypeptide for purposes herein is thus determined by such considerations.

As a general proposition, the total pharmaceutically effective amount of CK α -5 polypeptide administered parenterally per dose will be in the range of about 1 μ g/kg/day to 10 mg/kg/day of patient body weight, although, as noted above, this will be subject to therapeutic discretion. More preferably, this dose is at least 0.01 mg/kg/day, and most preferably for humans between about 0.01 and 1 mg/kg/day for the hormone. If given continuously, the CK α -5 polypeptide is typically administered at a dose rate of about 1 μ g/kg/hour to about 50 μ g/kg/hour, either by 1-4 injections per day or by continuous subcutaneous infusions, for example, using a mini-pump. An intravenous bag solution may also be employed. The length of

treatment needed to observe changes and the interval following treatment for responses to occur appears to vary depending on the desired effect.

Pharmaceutical compositions containing the CK α -5 of the invention may be administered orally, rectally, parenterally, intracisternally, intravaginally, intraperitoneally, topically (as by
5 powders, ointments, drops or transdermal patch), buccally, or as an oral or nasal spray. By "pharmaceutically acceptable carrier" is meant a non-toxic solid, semisolid or liquid filler, diluent, encapsulating material or formulation auxiliary of any type. The term "parenteral" as used herein refers to modes of administration which include intravenous, intramuscular, intraperitoneal, intrasternal, subcutaneous and intraarticular injection and infusion.

10 The CK α -5 polypeptide is also suitably administered by sustained-release systems. Suitable examples of sustained-release compositions include semi-permeable polymer matrices in the form of shaped articles, e.g., films, or microcapsules. Sustained-release matrices include polylactides (U.S. Pat. No. 3,773,919, EP 58,481), copolymers of L-glutamic acid and gamma-ethyl-L-glutamate (Sidman, U. et al., *Biopolymers* 22:547-556 (1983)), poly (2- hydroxyethyl
15 methacrylate) (R. Langer et al., *J. Biomed. Mater. Res.* 15:167-277 (1981), and R. Langer, *Chem. Tech.* 12:98-105 (1982)), ethylene vinyl acetate (R. Langer et al., Id.) or poly-D- (-)-3-hydroxybutyric acid (EP 133,988). Sustained-release CK α -5 polypeptide compositions also include liposomally entrapped CK α -5 polypeptide. Liposomes containing CK α -5 polypeptide are prepared by methods known per se: DE 3,218,121; Epstein et al., *Proc. Natl. Acad. Sci.*
20 (USA) 82:3688-3692 (1985); Hwang et al., *Proc. Natl. Acad. Sci. (USA)* 77:4030-4034 (1980); EP 52,322; EP 36,676; EP 88,046; EP 143,949; EP 142,641; Japanese Pat. Appl. 83-118008; U.S. Pat. Nos. 4,485,045 and 4,544,545; and EP 102,324. Ordinarily, the liposomes are of the small (about 200-800 Angstroms) unilamellar type in which the lipid content is greater than about 30 mol. percent cholesterol, the selected proportion being adjusted for the optimal CK α -5
25 polypeptide therapy.

For parenteral administration, in one embodiment, the CK α -5 polypeptide is formulated generally by mixing it at the desired degree of purity, in a unit dosage injectable form (solution, suspension, or emulsion), with a pharmaceutically acceptable carrier, i.e., one that is non-toxic to
30 recipients at the dosages and concentrations employed and is compatible with other ingredients of the formulation. For example, the formulation preferably does not include oxidizing agents and other compounds that are known to be deleterious to polypeptides.

Generally, the formulations are prepared by contacting the CK α -5 polypeptide uniformly and intimately with liquid carriers or finely divided solid carriers or both. Then, if necessary, the product is shaped into the desired formulation. Preferably the carrier is a parenteral carrier, more
35 preferably a solution that is isotonic with the blood of the recipient. Examples of such carrier

vehicles include water, saline, Ringer's solution, and dextrose solution. Non-aqueous vehicles such as fixed oils and ethyl oleate are also useful herein, as well as liposomes.

The carrier suitably contains minor amounts of additives such as substances that enhance isotonicity and chemical stability. Such materials are non-toxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, succinate, acetic acid, and other organic acids or their salts; antioxidants such as ascorbic acid; low molecular weight (less than about ten residues) polypeptides, e.g., polyarginine or tripeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids, such as glycine, glutamic acid, aspartic acid, or arginine; monosaccharides, disaccharides, and other carbohydrates including cellulose or its derivatives, glucose, manose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; counterions such as sodium; and/or nonionic surfactants such as polysorbates, poloxamers, or PEG.

The CK α -5 polypeptide is typically formulated in such vehicles at a concentration of about 0.1 mg/ml to 100 mg/ml, preferably 1-10 mg/ml, at a pH of about 3 to 8. It will be understood that the use of certain of the foregoing excipients, carriers, or stabilizers will result in the formation of CK α -5 polypeptide salts.

CK α -5 polypeptide to be used for therapeutic administration must be sterile. Sterility is readily accomplished by filtration through sterile filtration membranes (e.g., 0.2 micron membranes). Therapeutic CK α -5 polypeptide compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

CK α -5 polypeptide ordinarily will be stored in unit or multi-dose containers, for example, sealed ampoules or vials, as an aqueous solution or as a lyophilized formulation for reconstitution. As an example of a lyophilized formulation, 10-ml vials are filled with 5 ml of sterile-filtered 1% (w/v) aqueous CK α -5 polypeptide solution, and the resulting mixture is lyophilized. The infusion solution is prepared by reconstituting the lyophilized CK α -5 polypeptide using bacteriostatic Water-for-Injection.

The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention. Associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration. In addition, the polypeptides of the present invention may be employed in conjunction with other therapeutic compounds.

Agonists and Antagonists - Assays and Molecules

The invention also provides a method of screening compounds to identify those which enhance or block the action of CK α -5 on cells, such as its interaction with CK α -5-binding
5 molecules such as receptor molecules. An agonist is a compound which increases the natural biological functions of CK α -5 or which functions in a manner similar to CK α -5, while antagonists decrease or eliminate such functions.

In another aspect of this embodiment the invention provides a method for identifying a receptor protein or other ligand-binding protein which binds specifically to a CK α -5 polypeptide.

10 For example, a cellular compartment, such as a membrane or a preparation thereof, may be prepared from a cell that expresses a molecule that binds CK α -5. The preparation is incubated with labeled CK α -5 and complexes of CK α -5 bound to the receptor or other binding protein are isolated and characterized according to routine methods known in the art.

Alternatively, the CK α -5 polypeptide may be bound to a solid support so that binding molecules
15 solubilized from cells are bound to the column and then eluted and characterized according to routine methods.

In the assay of the invention for agonists or antagonists, a cellular compartment, such as a membrane or a preparation thereof, may be prepared from a cell that expresses a molecule that binds CK α -5, such as a molecule of a signaling or regulatory pathway modulated by CK α -5.

20 The preparation is incubated with labeled CK α -5 in the absence or the presence of a candidate molecule which may be a CK α -5 agonist or antagonist. The ability of the candidate molecule to bind the binding molecule is reflected in decreased binding of the labeled ligand. Molecules which bind gratuitously, i.e., without inducing the effects of CK α -5 on binding the CK α -5 binding molecule, are most likely to be good antagonists. Molecules that bind well and elicit
25 effects that are the same as or closely related to CK α -5 are agonists.

CK α -5-like effects of potential agonists and antagonists may be measured, for instance, by determining activity of a second messenger system following interaction of the candidate molecule with a cell or appropriate cell preparation, and comparing the effect with that of CK α -5 or molecules that elicit the same effects as CK α -5. Second messenger systems that may be useful
30 in this regard include but are not limited to AMP guanylate cyclase, ion channel or phosphoinositide hydrolysis second messenger systems.

Another example of an assay for CK α -5 antagonists is a competitive assay that combines CK α -5 and a potential antagonist with membrane-bound CK α -5 receptor molecules or

recombinant CK α -5 receptor molecules under appropriate conditions for a competitive inhibition assay. CK α -5 can be labeled, such as by radioactivity, such that the number of CK α -5 molecules bound to a receptor molecule can be determined accurately to assess the effectiveness of the potential antagonist.

- 5 Potential antagonists include small organic molecules, peptides, polypeptides and antibodies that bind to a polypeptide of the invention and thereby inhibit or extinguish its activity. Potential antagonists also may be small organic molecules, a peptide, a polypeptide such as a closely related protein or antibody that binds the same sites on a binding molecule, such as a receptor molecule, without inducing CK α -5-induced activities, thereby preventing the action of
- 10 CK α -5 by excluding CK α -5 from binding.

- Other potential antagonists include antisense molecules. Antisense technology can be used to control gene expression through antisense DNA or RNA or through triple-helix formation. Antisense techniques are discussed, for example, in Okano, *J. Neurochem.* 56: 560 (1991); "Oligodeoxynucleotides as Antisense Inhibitors of Gene Expression." CRC Press, Boca
- 15 Raton, FL (1988). Triple helix formation is discussed in, for instance *Lee et al., Nucleic Acids Research* 6: 3073 (1979); Cooney *et al., Science* 241: 456 (1988); and *Dervan et al., Science* 251: 1360 (1991). The methods are based on binding of a polynucleotide to a complementary DNA or RNA. For example, the 5' coding portion of a polynucleotide that encodes the mature
- 20 polypeptide of the present invention may be used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription thereby preventing transcription and the production of CK α -5. The antisense RNA oligonucleotide hybridizes to the mRNA *in vivo* and blocks translation of the mRNA molecule into CK α -5 polypeptide. The oligonucleotides described above can also be delivered to cells such that the antisense RNA or DNA may be
- 25 expressed *in vivo* to inhibit production of CK α -5 protein.

The agonists and antagonists may be employed in a composition with a pharmaceutically acceptable carrier, e.g., as described above.

- The antagonists may be employed for instance to inhibit the chemotaxis and activation of macrophages and their precursors, and of neutrophils, basophils, B lymphocytes and some T-cell
- 30 subsets, e.g., activated and CD8 cytotoxic T cells and natural killer cells, in certain auto-immune and chronic inflammatory and infective diseases. Examples of auto-immune diseases include multiple sclerosis, and insulin-dependent diabetes. The antagonists may also be employed to treat infectious diseases including silicosis, sarcoidosis, idiopathic pulmonary fibrosis by preventing the recruitment and activation of mononuclear phagocytes. They may also be employed to treat
- 35 idiopathic hyper-eosinophilic syndrome by preventing eosinophil production and migration. Endotoxic shock may also be treated by the antagonists by preventing the migration of

macrophages and their production of the human chemokine polypeptides of the present invention. The antagonists may also be employed for treating atherosclerosis, by preventing monocyte infiltration in the artery wall. The antagonists may also be employed to treat histamine-mediated allergic reactions and immunological disorders including late phase allergic reactions, chronic urticaria, and atopic dermatitis by inhibiting chemokine-induced mast cell and basophil degranulation and release of histamine. IgE-mediated allergic reactions such as allergic asthma, rhinitis, and eczema may also be treated. The antagonists may also be employed to treat chronic and acute inflammation by preventing the attraction of monocytes to a wound area. They may also be employed to regulate normal pulmonary macrophage populations, since chronic and acute inflammatory pulmonary diseases are associated with sequestration of mononuclear phagocytes in the lung. Antagonists may also be employed to treat rheumatoid arthritis by preventing the attraction of monocytes into synovial fluid in the joints of patients. Monocyte influx and activation plays a significant role in the pathogenesis of both degenerative and inflammatory arthropathies. The antagonists may be employed to interfere with the deleterious cascades attributed primarily to IL-1 and TNF, which prevents the biosynthesis of other inflammatory cytokines. In this way, the antagonists may be employed to prevent inflammation. The antagonists may also be employed to inhibit prostaglandin-independent fever induced by chemokines. The antagonists may also be employed to treat cases of bone marrow failure, for example, aplastic anemia and myelodysplastic syndrome. The antagonists may also be employed to treat asthma and allergy by preventing eosinophil accumulation in the lung. The antagonists may also be employed to treat subepithelial basement membrane fibrosis which is a prominent feature of the asthmatic lung. Antibodies against CK α -5 may be employed to bind to and inhibit CK α -5 activity to treat ARDS, by preventing infiltration of neutrophils into the lung after injury. Any of the above antagonists may be employed in a composition with a pharmaceutically acceptable carrier, e.g., as hereinafter described.

Gene Mapping

The nucleic acid molecules of the present invention are also valuable for chromosome identification. The sequence is specifically targeted to and can hybridize with a particular location on an individual human chromosome. Moreover, there is a current need for identifying particular sites on the chromosome. Few chromosome marking reagents based on actual sequence data (repeat polymorphisms) are presently available for marking chromosomal location. The mapping of DNAs to chromosomes according to the present invention is an important first step in correlating those sequences with genes associated with disease.

In certain preferred embodiments in this regard, the cDNA herein disclosed is used to clone genomic DNA of a CK α -5 protein gene. This can be accomplished using a variety of well known techniques and libraries, which generally are available commercially. The genomic DNA then is used for *in situ* chromosome mapping using well known techniques for this purpose.

In addition, in some cases, sequences can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp) from the cDNA. Computer analysis of the 3' untranslated region of the gene is used to rapidly select primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers are then used for PCR
5 screening of somatic cell hybrids containing individual human chromosomes. Fluorescence *in situ* hybridization ("FISH") of a cDNA clone to a metaphase chromosomal spread can be used to provide a precise chromosomal location in one step. This technique can be used with probes from the cDNA as short as 50 or 60 bp. For a review of this technique, see Verma *et al.*, *Human Chromosomes: A Manual Of Basic Techniques*, Pergamon Press, New York (1988).

10 Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. Such data are found, for example, in V. McKusick, *Mendelian Inheritance In Man*, available on-line through Johns Hopkins University, Welch Medical Library. The relationship between genes and diseases that have been mapped to the same chromosomal region are then identified through
15 linkage analysis (coinheritance of physically adjacent genes).

Next, it is necessary to determine the differences in the cDNA or genomic sequence between affected and unaffected individuals. If a mutation is observed in some or all of the affected individuals but not in any normal individuals, then the mutation is likely to be the causative agent of the disease.

20 Having generally described the invention, the same will be more readily understood by reference to the following examples, which are provided by way of illustration and are not intended as limiting.

Examples

Example 1(a): Expression and Purification of "His-tagged" CK α -5 in *E. coli*

25 The bacterial expression vector pQE60 is used for bacterial expression in this example. (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311). pQE60 encodes ampicillin antibiotic resistance ("Ampr") and contains a bacterial origin of replication ("ori"), an IPTG inducible promoter, a ribosome binding site ("RBS"), six codons encoding histidine residues that allow affinity purification using nickel-nitrilo-tri-acetic acid ("Ni-NTA") affinity resin sold by
30 QIAGEN, Inc., *supra*, and suitable single restriction enzyme cleavage sites. These elements are arranged such that an inserted DNA fragment encoding a polypeptide expresses that polypeptide with the six His residues (i.e., a "6 X His tag") covalently linked to the carboxyl terminus of that polypeptide.

The DNA sequence encoding the desired portion CK α -5 protein lacking the hydrophobic
35 leader sequence and transmembrane domain is amplified from the deposited cDNA clone using PCR oligonucleotide primers which anneal to the amino terminal sequences of the desired portion

of the CK α -5 protein and to sequences in the deposited construct 3' to the cDNA coding sequence. Additional nucleotides containing restriction sites to facilitate cloning in the pQE60 vector are added to the 5' and 3' sequences, respectively.

For cloning the mature protein, the 5' primer has the sequence

5' GCGCCATGGAATGGCAACGAGGGCAGC 3' (SEQ ID NO: 15) containing the underlined NcoI restriction site. One of ordinary skill in the art would appreciate, of course, that the point in the protein coding sequence where the 5' primer begins may be varied to amplify a DNA segment encoding any desired portion of the complete protein shorter or longer than the mature form. The 3' primer has the sequence 5' CGCAAGCTTTTATGTGGCTGATGTCCTGGC 3' (SEQ ID NO: 16) containing the underlined HindIII restriction.

The amplified CK α -5 DNA fragment and the vector pQE60 are digested with NcoI and HindIII and the digested DNAs are then ligated together. Insertion of the CK α -5 DNA into the restricted pQE60 vector places the CK α -5 protein coding region downstream from the IPTG-inducible promoter and in-frame with an initiating AUG and the six histidine codons.

Alternatively, a preferred bacterial expression vector "pHE4-5" containing an ampicillin resistance gene may be used in this example. pHE4-5/MPIFD23 vector plasmid DNA contains a filler insert between unique restriction enzyme sites *Nde*I and *Asp*718 and was deposited with the American Type Culture Collection, 12301 Park Lawn Drive, Rockville, Maryland 20852, on September 30, 1997 and given Accession No. 209311. Using 5' and 3' primers described herein with restriction enzyme sites for *Nde*I and *Asp* 718 substituted for the *Nco*I and *Hind*III sites in the respective primers, a suitable CK α -5 encoding DNA fragment for subcloning into pHE4-5 can be amplified. The stuffer DNA insert in pHE4-5/MPIFD23 should be removed prior to ligating the CK α -5 fragment to pHE4-5. pHE4-5 contains a strong bacterial promoter allowing for high yields of most heterologous proteins.

The ligation mixture is transformed into competent *E. coli* cells using standard procedures such as those described in Sambrook et al., *Molecular Cloning: a Laboratory Manual, 2nd Ed.*; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989). *E. coli* strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses the lac repressor and confers kanamycin resistance ("Kanr"), is used in carrying out the illustrative example described herein. This strain, which is only one of many that are suitable for expressing CK α -5 protein, is available commercially from QIAGEN, Inc., *supra*. Transformants are identified by their ability to grow on LB plates in the presence of ampicillin and kanamycin. Plasmid DNA is isolated from resistant colonies and the identity of the cloned DNA confirmed by restriction analysis, PCR and DNA sequencing.

Clones containing the desired constructs are grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 µg/ml) and kanamycin (25 µg/ml). The O/N culture is used to inoculate a large culture, at a dilution of approximately 1:25 to 1:250. The cells are grown to an optical density at 600 nm ("OD600") of between 0.4 and 0.6. Isopropyl-β-D-thiogalactopyranoside ("IPTG") is then added to a final concentration of 1 mM to induce transcription from the lac repressor sensitive promoter, by inactivating the lacI repressor. Cells subsequently are incubated further for 3 to 4 hours. Cells then are harvested by centrifugation.

The cells are then stirred for 3-4 hours at 4° C in 6M guanidine-HCl, pH 8. The cell debris is removed by centrifugation, and the supernatant containing the CKα-5 is loaded onto a nickel-nitrilo-tri-acetic acid ("Ni-NTA") affinity resin column (available from QIAGEN, Inc., *supra*). Proteins with a 6 x His tag bind to the Ni-NTA resin with high affinity and can be purified in a simple one-step procedure (for details see: The QIAexpressionist, 1995, QIAGEN, Inc., *supra*). Briefly the supernatant is loaded onto the column in 6 M guanidine-HCl, pH 8, the column is first washed with 10 volumes of 6 M guanidine-HCl, pH 8, then washed with 10 volumes of 6 M guanidine-HCl pH 6, and finally the CKα-5 is eluted with 6 M guanidine-HCl, pH 5.

The purified protein is then renatured by dialyzing it against phosphate-buffered saline (PBS) or 50 mM Na-acetate, pH 6 buffer plus 200 mM NaCl. Alternatively, the protein can be successfully refolded while immobilized on the Ni-NTA column. The recommended conditions are as follows: renature using a linear 6M-1M urea gradient in 500 mM NaCl, 20% glycerol, 20 mM Tris/HCl pH 7.4, containing protease inhibitors. The renaturation should be performed over a period of 1.5 hours or more. After renaturation the proteins can be eluted by the addition of 250 mM imidazole. Imidazole is removed by a final dialyzing step against PBS or 50 mM sodium acetate pH 6 buffer plus 200 mM NaCl. The purified protein is stored at 4° C or frozen at -80° C.

The following alternative method may be used to purify CKα-5 expressed in *E. coli* when it is present in the form of inclusion bodies. Unless otherwise specified, all of the following steps are conducted at 4-10°C.

Upon completion of the production phase of the *E. coli* fermentation, the cell culture is cooled to 4-10°C and the cells are harvested by continuous centrifugation at 15,000 rpm (Heraeus Sepatech). On the basis of the expected yield of protein per unit weight of cell paste and the amount of purified protein required, an appropriate amount of cell paste, by weight, is suspended in a buffer solution containing 100 mM Tris, 50 mM EDTA, pH 7.4. The cells are dispersed to a homogeneous suspension using a high shear mixer.

The cells were then lysed by passing the solution through a microfluidizer (Microfluidics, Corp. or APV Gaulin, Inc.) twice at 4000-6000 psi. The homogenate is then mixed with NaCl

solution to a final concentration of 0.5 M NaCl, followed by centrifugation at 7000 xg for 15 min. The resultant pellet is washed again using 0.5M NaCl, 100 mM Tris, 50 mM EDTA, pH 7.4.

The resulting washed inclusion bodies are solubilized with 1.5 M guanidine hydrochloride (GuHCl) for 2-4 hours. After 7000 xg centrifugation for 15 min., the pellet is discarded and the CK α -5 polypeptide-containing supernatant is incubated at 4°C overnight to allow further GuHCl extraction.

Following high speed centrifugation (30,000 xg) to remove insoluble particles, the GuHCl solubilized protein is refolded by quickly mixing the GuHCl extract with 20 volumes of buffer containing 50 mM sodium, pH 4.5, 150 mM NaCl, 2 mM EDTA by vigorous stirring. The refolded diluted protein solution is kept at 4°C without mixing for 12 hours prior to further purification steps.

To clarify the refolded CK α -5 polypeptide solution, a previously prepared tangential filtration unit equipped with 0.16 μ m membrane filter with appropriate surface area (e.g., Filtron), equilibrated with 40 mM sodium acetate, pH 6.0 is employed. The filtered sample is loaded onto a cation exchange resin (e.g., Poros HS-50, Perseptive Biosystems). The column is washed with 40 mM sodium acetate, pH 6.0 and eluted with 250 mM, 500 mM, 1000 mM, and 1500 mM NaCl in the same buffer, in a stepwise manner. The absorbance at 280 nm of the effluent is continuously monitored. Fractions are collected and further analyzed by SDS-PAGE.

Fractions containing the CK α -5 polypeptide are then pooled and mixed with 4 volumes of water. The diluted sample is then loaded onto a previously prepared set of tandem columns of strong anion (Poros HQ-50, Perseptive Biosystems) and weak anion (Poros CM-20, Perseptive Biosystems) exchange resins. The columns are equilibrated with 40 mM sodium acetate, pH 6.0. Both columns are washed with 40 mM sodium acetate, pH 6.0, 200 mM NaCl. The CM-20 column is then eluted using a 10 column volume linear gradient ranging from 0.2 M NaCl, 50 mM sodium acetate, pH 6.0 to 1.0 M NaCl, 50 mM sodium acetate, pH 6.5. Fractions are collected under constant A_{280} monitoring of the effluent. Fractions containing the CK α -5 polypeptide (determined, for instance, by 16% SDS-PAGE) are then pooled.

The resultant CK α -5 polypeptide exhibits greater than 95% purity after the above refolding and purification steps. No major contaminant bands are observed from Commassie blue stained 16% SDS-PAGE gel when 5 μ g of purified protein is loaded. The purified protein is also tested for endotoxin/LPS contamination, and typically the LPS content is less than 0.1 ng/ml according to LAL assays.

Example 2: Cloning and Expression of CK α -5 protein in a Baculovirus

Expression System

In this illustrative example, the plasmid shuttle vector pA2 is used to insert the cloned DNA encoding complete protein, including its naturally associated secretory signal (leader) sequence, into a baculovirus to express the mature CK α -5 protein, using standard methods as described in Summers *et al.*, *A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures*, Texas Agricultural Experimental Station Bulletin No. 1555 (1987). This expression vector contains the strong polyhedrin promoter of the *Autographa californica* nuclear polyhedrosis virus (AcMNPV) followed by convenient restriction sites such as BamHI, Xba I and Asp718. The polyadenylation site of the simian virus 40 ("SV40") is used for efficient polyadenylation. For easy selection of recombinant virus, the plasmid contains the beta-galactosidase gene from *E. coli* under control of a weak *Drosophila* promoter in the same orientation, followed by the polyadenylation signal of the polyhedrin gene. The inserted genes are flanked on both sides by viral sequences for cell-mediated homologous recombination with wild-type viral DNA to generate a viable virus that express the cloned polynucleotide.

Many other baculovirus vectors could be used in place of the vector above, such as pAc373, pVL941 and pAcIM1, as one skilled in the art would readily appreciate, as long as the construct provides appropriately located signals for transcription, translation, secretion and the like, including a signal peptide and an in-frame AUG as required. Such vectors are described, for instance, in Luckow *et al.*, *Virology* 170:31-39 (1989).

The cDNA sequence encoding the full length CK α -5 protein in the deposited clone, including the AUG initiation codon and the naturally associated leader sequence shown in SEQ ID NO:2, is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the gene. The 5' primer has the sequence 5' CGCGGATCCGCCATCATGGGACGGGACTTGCGG 3' (SEQ ID NO:17) containing the underlined BamHI restriction enzyme site, an efficient signal for initiation of translation in eukaryotic cells, as described by Kozak, M., *J. Mol. Biol.* 196:947-950 (1987). The 3' primer has the sequence 5' GCGTCTAGATCAGGTATTAGAGTCAGG 3' (SEQ ID NO:18) containing the underlined XbaI restriction site. Those of ordinary skill in the art will recognize that other primers could be used to generate nucleic acid fragments encoding shorter polypeptides. For example, the 5' primer above (SEQ ID NO:19) with the following 3' primer could be used to amplify the nucleic acid fragment encoding the soluble extracellular domain:

5' GCGTCTAGATTATGTGGCTGATGTCCTGGC 3' (SEQ ID NO:19). The 3' primer contains the underlined XbaI site.

The amplified fragment is isolated from a 1% agarose gel using a commercially available kit ("GeneClean," BIO 101 Inc., La Jolla, Ca.). The fragment then is digested with BamHI and XbaI and again is purified on a 1% agarose gel.

The plasmid is digested with the restriction enzymes BamHI and XbaI and optionally, can be dephosphorylated using calf intestinal phosphatase, using routine procedures known in the art. The DNA is then isolated from a 1% agarose gel using a commercially available kit ("GeneClean" BIO 101 Inc., La Jolla, Ca.).

Fragment and the dephosphorylated plasmid are ligated together with T4 DNA ligase. *E. coli* HB101 or other suitable *E. coli* hosts such as XL-1 Blue (Statagene Cloning Systems, La Jolla, CA) cells are transformed with the ligation mixture and spread on culture plates. Bacteria are identified that contain the plasmid with the human CK α -5 gene by digesting DNA from individual colonies using BamHI and XbaI and then analyzing the digestion product by gel electrophoresis. The sequence of the cloned fragment is confirmed by DNA sequencing. This plasmid is designated herein pA2GPCK α -5.

Five μ g of the plasmid pA2GPCK α -5 is co-transfected with 1.0 μ g of a commercially available linearized baculovirus DNA ("BaculoGold™ baculovirus DNA", Pharmingen, San Diego, CA), using the lipofection method described by Felgner et al., *Proc. Natl. Acad. Sci. USA* 84: 7413-7417 (1987). One μ g of BaculoGold™ virus DNA and 5 μ g of the plasmid pA2GPCK α -5 are mixed in a sterile well of a microtiter plate containing 50 μ l of serum-free Grace's medium (Life Technologies Inc., Gaithersburg, MD). Afterwards, 10 μ l Lipofectin plus 90 μ l Grace's medium are added, mixed and incubated for 15 minutes at room temperature. Then the transfection mixture is added drop-wise to Sf9 insect cells (ATCC CRL 1711) seeded in a 35 mm tissue culture plate with 1 ml Grace's medium without serum. The plate is then incubated for 5 hours at 27° C. The transfection solution is then removed from the plate and 1 ml of Grace's insect medium supplemented with 10% fetal calf serum is added. Cultivation is then continued at 27° C for four days.

After four days the supernatant is collected and a plaque assay is performed, as described by Summers and Smith, *supra*. An agarose gel with "Blue Gal" (Life Technologies Inc., Gaithersburg) is used to allow easy identification and isolation of gal-expressing clones, which produce blue-stained plaques. (A detailed description of a "plaque assay" of this type can also be found in the user's guide for insect cell culture and baculovirology distributed by Life Technologies Inc., Gaithersburg, page 9-10). After appropriate incubation, blue stained plaques are picked with the tip of a micropipettor (e.g., Eppendorf). The agar containing the recombinant viruses is then resuspended in a microcentrifuge tube containing 200 μ l of Grace's medium and the suspension containing the recombinant baculovirus is used to infect Sf9 cells seeded in 35 mm

bacteria are identified that contain the fragment inserted into plasmid pC4 using, for instance, restriction enzyme analysis.

Chinese hamster ovary cells lacking an active DHFR gene are used for transfection. Five μ g of the expression plasmid pC4 is cotransfected with 0.5 μ g of the plasmid pSVneo using lipofectin (Felgner et al., *supra*). The plasmid pSV2-neo contains a dominant selectable marker, the *neo* gene from Tn5 encoding an enzyme that confers resistance to a group of antibiotics including G418. The cells are seeded in alpha minus MEM supplemented with 1 mg/ml G418. After 2 days, the cells are trypsinized and seeded in hybridoma cloning plates (Greiner, Germany) in alpha minus MEM supplemented with 10, 25, or 50 ng/ml of methotrexate plus 1 mg/ml G418. After about 10-14 days single clones are trypsinized and then seeded in 6-well petri dishes or 10 ml flasks using different concentrations of methotrexate (50 nM, 100 nM, 200 nM, 400 nM, 800 nM). Clones growing at the highest concentrations of methotrexate are then transferred to new 6-well plates containing even higher concentrations of methotrexate (1 μ M, 2 μ M, 5 μ M, 10 mM, 20 mM). The same procedure is repeated until clones are obtained which grow at a concentration of 100 - 200 μ M. Expression of the desired gene product is analyzed, for instance, by SDS-PAGE and Western blot or by reversed phase HPLC analysis.

Example 4: Tissue distribution of CK α -5 mRNA expression

Northern blot analysis is carried out to examine CK α -5 gene expression in human tissues, using methods described by, among others, Sambrook *et al.*, cited above. A cDNA probe containing the entire nucleotide sequence of the CK α -5 protein (SEQ ID NO:1) is labeled with 32 P using the *rediprime*TM DNA labeling system (Amersham Life Science), according to manufacturer's instructions. After labeling, the probe is purified using a CHROMA SPIN-100TM column (Clontech Laboratories, Inc.), according to manufacturer's protocol number PT1200-1. The purified labeled probe is then used to examine various human tissues for CK α -5 mRNA.

Multiple Tissue Northern (MTN) blots containing various human tissues (H) or human immune system tissues (IM) are obtained from Clontech and are examined with the labeled probe using *ExpressHyb*TM hybridization solution (Clontech) according to manufacturer's protocol number PT1190-1. Following hybridization and washing, the blots are mounted and exposed to film at -70° C overnight, and films developed according to standard procedures.

It will be clear that the invention may be practiced otherwise than as particularly described in the foregoing description and examples. Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, are within the scope of the appended claims.

The entire disclosure of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein are hereby incorporated by reference.

Further, the sequence listing in both hard copy and electronic form submitted herewith is incorporated herein by reference.

marker is the enzyme glutamine synthase (GS) (Murphy et al., *Biochem J.* 227:277-279 (1991); Bebbington et al., *Bio/Technology* 10:169-175 (1992)). Using these markers, the mammalian cells are grown in selective medium and the cells with the highest resistance are selected. These cell lines contain the amplified gene(s) integrated into a chromosome. Chinese hamster ovary (CHO) and NSO cells are often used for the production of proteins.

The expression vectors pC1 and pC4 contain the strong promoter (LTR) of the Rous Sarcoma Virus (Cullen et al., *Molecular and Cellular Biology*, 438-447 (March, 1985)) plus a fragment of the CMV-enhancer (Boshart et al., *Cell* 41:521-530 (1985)). Multiple cloning sites, e.g., with the restriction enzyme cleavage sites BamHI, XbaI and Asp718, facilitate the cloning of the gene of interest. The vectors contain in addition the 3' intron, the polyadenylation and termination signal of the rat preproinsulin gene.

Example 3(a): Cloning and Expression in COS Cells

The expression plasmid, pCK α -5HA, is made by cloning a portion of the cDNA encoding the extracellular domain of the CK α -5 protein into the expression vector pcDNAI/Amp or pcDNAIII (which can be obtained from Invitrogen, Inc.).

The expression vector pcDNAI/amp contains: (1) an *E. coli* origin of replication effective for propagation in *E. coli* and other prokaryotic cells; (2) an ampicillin resistance gene for selection of plasmid-containing prokaryotic cells; (3) an SV40 origin of replication for propagation in eukaryotic cells; (4) a CMV promoter, a polylinker, an SV40 intron; (5) several codons encoding a hemagglutinin fragment (i.e., an "HA" tag to facilitate purification) followed by a termination codon and polyadenylation signal arranged so that a cDNA can be conveniently placed under expression control of the CMV promoter and operably linked to the SV40 intron and the polyadenylation signal by means of restriction sites in the polylinker. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein described by Wilson et al., *Cell* 37: 767 (1984). The fusion of the HA tag to the target protein allows easy detection and recovery of the recombinant protein with an antibody that recognizes the HA epitope. pcDNAIII contains, in addition, the selectable neomycin marker.

A DNA fragment encoding the soluble extracellular domain of the CK α -5 polypeptide is cloned into the polylinker region of the vector so that recombinant protein expression is directed by the CMV promoter. The plasmid construction strategy is as follows. The CK α -5 cDNA of the deposited clone is amplified using primers that contain convenient restriction sites, much as described above for construction of vectors for expression of CK α -5 in *E. coli*. Suitable primers include the following, which are used in this example. The 5' primer, containing the underlined BamHI site, a Kozak sequence, and an AUG start codon, has the following sequence:
5' CGCGGATCCGCCATCATGGGACGGGACTTGCGG 3' (SEQ ID NO:17). The 3'

primer, containing the underlined XbaI site has the following sequence:

5' GCGTCTAGATCAGGTATTAGAGTCAGG 3' (SEQ ID NO:19).

The PCR amplified DNA fragment and the vector, pcDNAI/Amp, are digested with BamHI and XbaI and then ligated. The ligation mixture is transformed into *E. coli* strain SURE (available from Stratagene Cloning Systems, 11099 North Torrey Pines Road, La Jolla, CA 92037), and the transformed culture is plated on ampicillin media plates which then are incubated to allow growth of ampicillin resistant colonies. Plasmid DNA is isolated from resistant colonies and examined by restriction analysis or other means for the presence of the fragment encoding the extracellular domain of the CK α -5 polypeptide

For expression of recombinant CK α -5, COS cells are transfected with an expression vector, as described above, using DEAE-DEXTRAN, as described, for instance, in Sambrook et al., *Molecular Cloning: a Laboratory Manual*, Cold Spring Laboratory Press, Cold Spring Harbor, New York (1989). Cells are incubated under conditions for expression of CK α -5 by the vector.

Expression of the CK α -5-HA fusion protein is detected by radiolabeling and immunoprecipitation, using methods described in, for example Harlow et al., *Antibodies: A Laboratory Manual, 2nd Ed.*; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988). To this end, two days after transfection, the cells are labeled by incubation in media containing ^{35}S -cysteine for 8 hours. The cells and the media are collected, and the cells are washed and the lysed with detergent-containing RIPA buffer: 150 mM NaCl, 1% NP-40, 0.1% SDS, 1% NP-40, 0.5% DOC, 50 mM TRIS, pH 7.5, as described by Wilson et al. cited above. Proteins are precipitated from the cell lysate and from the culture media using an HA-specific monoclonal antibody. The precipitated proteins then are analyzed by SDS-PAGE and autoradiography. An expression product of the expected size is seen in the cell lysate, which is not seen in negative controls.

Example 3(b): Cloning and Expression in CHO Cells

The vector pC4 is used for the expression of CK α -5 polypeptide in this example. Plasmid pC4 is a derivative of the plasmid pSV2-dhfr (ATCC Accession No. 37146). The plasmid contains the mouse DHFR gene under control of the SV40 early promoter. Chinese hamster ovary- or other cells lacking dihydrofolate activity that are transfected with these plasmids can be selected by growing the cells in a selective medium (alpha minus MEM, Life Technologies) supplemented with the chemotherapeutic agent methotrexate. The amplification of the DHFR genes in cells resistant to methotrexate (MTX) has been well documented (see, e.g., Alt, F. W., Kellems, R. M., Bertino, J. R., and Schimke, R. T., 1978, *J. Biol. Chem.* 253:1357-1370, Hamlin, J. L. and Ma, C. 1990, *Biochem. et Biophys. Acta*, 1097:107-143,

Page, M. J. and Sydenham, M. A. 1991, *Biotechnology* 9:64-68). Cells grown in increasing concentrations of MTX develop resistance to the drug by overproducing the target enzyme, DHFR, as a result of amplification of the DHFR gene. If a second gene is linked to the DHFR gene, it is usually co-amplified and over-expressed. It is known in the art that this approach may be used to develop cell lines carrying more than 1,000 copies of the amplified gene(s). Subsequently, when the methotrexate is withdrawn, cell lines are obtained which contain the amplified gene integrated into one or more chromosome(s) of the host cell.

Plasmid pC4 contains for expressing the gene of interest the strong promoter of the long terminal repeat (LTR) of the Rouse Sarcoma Virus (Cullen, et al., *Molecular and Cellular Biology*, March 1985:438-447) plus a fragment isolated from the enhancer of the immediate early gene of human cytomegalovirus (CMV) (Boshart et al., *Cell* 41:521-530 (1985)). Downstream of the promoter are the following single restriction enzyme cleavage sites that allow the integration of the genes: BamHI, Xba I, and Asp718. Behind these cloning sites the plasmid contains the 3' intron and polyadenylation site of the rat preproinsulin gene. Other high efficiency promoters can also be used for the expression, e.g., the human β -actin promoter, the SV40 early or late promoters or the long terminal repeats from other retroviruses, e.g., HIV and HTLV. Clontech's Tet-Off and Tet-On gene expression systems and similar systems can be used to express the CK α -5 polypeptide in a regulated way in mammalian cells (Gossen, M., & Bujard, H. 1992, *Proc. Natl. Acad. Sci. USA* 89:5547-5551). For the polyadenylation of the mRNA other signals, e.g., from the human growth hormone or globin genes can be used as well. Stable cell lines carrying a gene of interest integrated into the chromosomes can also be selected upon co-transfection with a selectable marker such as gpt, G418 or hygromycin. It is advantageous to use more than one selectable marker in the beginning, e.g., G418 plus methotrexate.

The plasmid pC4 is digested with the restriction enzymes BamHI and XbaI and then dephosphorylated using calf intestinal phosphatase by procedures known in the art. The vector is then isolated from a 1% agarose gel.

The DNA sequence encoding the extracellular domain of the CK α -5 polypeptide is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the desired portion of the gene. The 5' and 3' primers are the same as those used in Example 3(a) above.

The amplified fragment is digested with the endonucleases BamHI and XbaI and then purified again on a 1% agarose gel. The isolated fragment and the dephosphorylated vector are then ligated with T4 DNA ligase. *E. coli* HB101 or XL-1 Blue cells are then transformed and bacteria are identified that contain the fragment inserted into plasmid pC4 using, for instance, restriction enzyme analysis.

Chinese hamster ovary cells lacking an active DHFR gene are used for transfection. Five μ g of the expression plasmid pC4 is cotransfected with 0.5 μ g of the plasmid pSVneo using

lipofectin (Felgner et al., *supra*). The plasmid pSV2-neo contains a dominant selectable marker, the *neo* gene from Tn5 encoding an enzyme that confers resistance to a group of antibiotics including G418. The cells are seeded in alpha minus MEM supplemented with 1 mg/ml G418. After 2 days, the cells are trypsinized and seeded in hybridoma cloning plates (Greiner, Germany) in alpha minus MEM supplemented with 10, 25, or 50 ng/ml of methotrexate plus 1 mg/ml G418. After about 10-14 days single clones are trypsinized and then seeded in 6-well petri dishes or 10 ml flasks using different concentrations of methotrexate (50 nM, 100 nM, 200 nM, 400 nM, 800 nM). Clones growing at the highest concentrations of methotrexate are then transferred to new 6-well plates containing even higher concentrations of methotrexate (1 μ M, 2 μ M, 5 μ M, 10 mM, 20 mM). The same procedure is repeated until clones are obtained which grow at a concentration of 100 - 200 μ M. Expression of the desired gene product is analyzed, for instance, by SDS-PAGE and Western blot or by reversed phase HPLC analysis.

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Multiple Tissue Northern (MTN) blots containing various human tissues (H) or human immune system tissues (IM) are obtained from Clontech and are examined with the labeled probe using ExpressHybTM hybridization solution (Clontech) according to manufacturer's protocol number PT1190-1. Following hybridization and washing, the blots are mounted and exposed to film at -70° C overnight, and films developed according to standard procedures.

It will be clear that the invention may be practiced otherwise than as particularly described in the foregoing description and examples. Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, are within the scope of the appended claims.

The entire disclosure of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein are hereby incorporated by reference.

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>2</u> , line <u>27</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input type="checkbox"/>	
Name of depositary institution American Type Culture Collection ("ATCC")	
Address of depositary institution (including postal code and country) 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	
Date of deposit 29 AUGUST 1997	Accession Number 209231
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

<p style="text-align: center;">For receiving Office use only</p> <p><input type="checkbox"/> This sheet was received with the international application</p> <p style="text-align: center;"> Elvira Rivera Paralegal Specialist INPD - PCT Operations (703) 305-3678 </p> <p>Authorized officer</p>	<p style="text-align: center;">For International Bureau use only</p> <p><input type="checkbox"/> This sheet was received by the International Bureau on:</p> <p>Authorized officer</p>
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What Is Claimed Is:

1. An isolated polynucleotide comprising a nucleotide sequence at least 95% identical to a sequence selected from the group consisting of:

- (a) a nucleotide sequence encoding residues -27 to 227 of SEQ ID NO:2;
- (b) a nucleotide sequence encoding residues -26 to 227 of SEQ ID NO:2;
- (c) a nucleotide sequence encoding residues 1-227 of SEQ ID NO:2;
- (d) a nucleotide sequence encoding residues 1-178 of SEQ ID NO:2;
- (e) a nucleotide sequence encoding the amino acid sequence of the full-length polypeptide encoded by the cDNA contained in ATCC Deposit No. 209231;

- (f) a nucleotide sequence encoding the amino acid sequence of the complete polypeptide, excepting the N-terminal methionine, encoded by the cDNA contained in ATCC Deposit No. 209231;

- (g) a nucleotide sequence encoding the amino acid sequence of the mature polypeptide encoded by the cDNA contained in ATCC Deposit No. 209231;

- (h) a nucleotide sequence encoding the amino acid sequence of the extracellular domain of the polypeptide encoded by the cDNA contained in ATCC Deposit No. 209231; and

- (i) a nucleotide sequence complementary to any of the nucleotide sequences in (a), (b), (c), (d), (e), (f), (g) or (h) above.

2. The polynucleotide of claim 1 wherein said polynucleotide comprises the nucleotide sequence shown as SEQ ID NO:1.

3. The polynucleotide of claim 1 wherein said polynucleotide has the portion of the nucleotide sequence in SEQ ID NO:1 which encodes residues -26 to 227 of SEQ ID NO:2.

4. The polynucleotide of claim 1 wherein said polynucleotide has the portion of the nucleotide sequence in SEQ ID NO:1 which encodes residues 1 to 178 of SEQ ID NO:2.

5. An isolated polynucleotide comprising a nucleotide sequence at least 95% identical to a sequence selected from the group consisting of:
- (a) a nucleotide sequence encoding a polypeptide comprising the amino acid sequence of residues n-178 of SEQ ID NO:2, where n is an integer in the range of -4 to 11;
 - (b) a nucleotide sequence encoding a polypeptide comprising the amino acid sequence of residues (-4)-m of SEQ ID NO:2, where m is an integer in the range of 55-178;
 - (c) a nucleotide sequence encoding a polypeptide having the amino acid sequence consisting of residues n-m of SEQ ID NO:2, where n and m are integers as defined respectively in (a) and (b) above;
 - (d) a nucleotide sequence encoding a polypeptide consisting of a portion of the complete CK α -5 amino acid sequence encoded by the cDNA contained in ATCC Deposit No. 209231 wherein said portion excludes from 23 to about 38 amino acids from the amino terminus of said complete amino acid sequence;
 - (e) a nucleotide sequence encoding a polypeptide consisting of a portion of the complete CK α -5 amino acid sequence encoded by the cDNA contained in ATCC Deposit No. 209231 wherein said portion excludes from 24 to about 172 amino acids from the carboxy terminus of said complete amino acid sequence; and
 - (f) a nucleotide sequence encoding a polypeptide consisting of a portion of the complete CK α -5 amino acid sequence encoded by the cDNA contained in ATCC Deposit No. 209231 wherein said portion includes a combination of any of the amino terminal and carboxy terminal deletions in (d) and (e), above.
6. The polynucleotide of claim 1 wherein said polynucleotide has the complete nucleotide sequence of the cDNA contained in ATCC Deposit No. 209231.
7. The polynucleotide of claim 1 wherein said polynucleotide has a nucleotide sequence encoding the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA contained in ATCC Deposit No. 209231.
8. The polynucleotide of claim 1 wherein said polynucleotide has the nucleotide sequence encoding the extracellular domain of the polypeptide encoded by the cDNA contained in ATCC Deposit No. 209231.
9. An polynucleotide comprising a polynucleotide which hybridizes under stringent hybridization conditions to a polynucleotide having a nucleotide sequence identical to a nucleotide sequence in (a), (b), (c), (d), (e), (f), (g), (h) or (i) of claim 1 wherein said polynucleotide which hybridizes does not hybridize under stringent hybridization conditions to a polynucleotide having a nucleotide sequence consisting of only A residues or of only T residues.

10. An isolated polynucleotide comprising a nucleotide sequence which encodes the amino acid sequence of an epitope-bearing portion of a CK α -5 polypeptide having an amino acid sequence in (a), (b), (c), (d), (e), (f), (g), (h) or (i) of claim 1.

11. The isolated polynucleotide of claim 10, which comprises a nucleotide sequence encoding an amino acid sequence selected from the group consisting of:

- (a) Tyr-12 to Ser-24 of SEQ ID NO:2;
- (b) Cys-55 to Val-63 in SEQ ID NO:2;
- (c) Thr-91 to Pro-110 in SEQ ID NO:2; and
- (d) Ser-120 to Ile-145 in SEQ ID NO:2.

12. A method for making a recombinant vector comprising inserting the isolated polynucleotide of claim 1 into a vector.

13. A recombinant vector produced by the method of claim 12.

14. A method of making a recombinant host cell comprising introducing the recombinant vector of claim 13 into a host cell.

15. A recombinant host cell produced by the method of claim 14.

16. A recombinant method for producing a CK α -5 polypeptide, comprising culturing the recombinant host cell of claim 15 under conditions such that said polypeptide is expressed and recovering said polypeptide.

17. An isolated CK α -5 polypeptide comprising an amino acid sequence at least 95% identical to an amino acid sequence selected from the group consisting of:

(a) the amino acid sequence shown as SEQ ID NO:2 or amino acid sequence encoded by the cDNA contained in the ATCC Deposit No. 209231;

(b) the amino acid shown as residues -26 to 227 of SEQ ID NO:2, or the complete amino acid sequence excepting the N-terminal methionine encoded by the cDNA contained in the ATCC Deposit No. 209231;

(c) the amino acid sequence shown as residues 1-227 of SEQ ID NO:2, or the amino acid sequence of the mature CK α -5 encoded by the cDNA contained in the ATCC Deposit No. 209231; and

(d) the amino acid sequence shown as residues 1-178 of SEQ ID NO:2; or the amino acid sequence of the extracellular domain of the polypeptide encoded by the cDNA contained in the ATCC Deposit No. 209231.

18. An isolated polypeptide encoded by the polynucleotide of claim 10.

19. An isolated antibody that binds specifically to the polypeptide of claim 18.

20. A method of treating an individual in need of a CK α -5 polypeptide comprising administering to the individual the isolated polypeptide of claim 17.

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FIGURE 1A

1.
CGGCGACTCTCTCCACCGGGCCGCGGGAGGCTCATGCAGCGCGGCTGGGTCCCGCGGC
61
GCCCCGATCGGGGAAGTGAAAGTGCCTCGGAGGAGGAGGGCCGGTCCGGCAGTGCAGCCG
121
CCTCACAGGTCTGGCGGACGGGCCAGGCGGGCGGCCTCCTGAACCGAACC GAATCGGCTCC
181
TCGGGCGGTCTCTCCCGCCCCCTCCTCGCCCGCCCGGAGTTTCTTTTCGGTTTCTTC
241
CAAGATTCTTGGCTTCCCTCGACGGAGCCGGGCCAGTGCGGGGGCGCAGGGCGCGGGA
301
GCTCCACCTCTCGGCTTTCCCTGCGTCCAGAGGCTGGCATGGCGGGGCCGAGTACTGA
361
GCGCACGGTCGGGGCACAGCAGGGCCGGTGGGTGCAGCTGGCTCGCGCCTCCTCTCCGGC
421
CGCCGTCTCTCCGGTCCCCGGCGAAAGCCATTGAGACACCAGCTGGACGTACGCGCCG
481
GAGCATGTCTGGGAGTCAGAGCGAGGTGGCTCCATCCCCGAGAGTCCGCGGAGCCCCGA
541
GATGGGACGGGACTTGCGGCCCCGGGTCCCGCGTGTCTCTGCTCCTGCTTCTGCTCCTGCT
M G R D L R P G S R V L L L L L L L L L 20
601
GGTGTAACCTGACTCAGCCAGGCAATGGCAACGAGGGCAGCGTCACTGGAAGTTGTTATTG
V Y L T O P G N G N E G S V T G S C Y C 40
661
TGGTAAAAGAATTTCTTCCGACTCCCCGCCATCGGTTTCAGTTCATGAATCGTCTCCGGAA
G K R I S S D S P P S V Q F M N R L R K 60
721
ACACCTGAGAGCTTACCATCGGTGTCTATACTACACGAGGTTCAGCTCCCTTCTCTGGAG
H L R A Y H R C L Y Y T R F Q L L S W S 80
781
CGTGTGTGGAGGCAACAAGGACCCATGGGTTCAGGAATTGATGAGCTGTCTTGATCTCAA
V C G G N K D P W V Q E L M S C L D L K 100

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FIGURE 1B

841

AGAATGTGGACATGCTTACTCGGGGATTGTGGCCCACCAGAAGCATTTACTTCCTACCAG
E C G H A Y S G I V A H Q K H L L P T S 120

901

CCCCCAACTTCTCAGGCCTCAGAGGGGGCATCTTCAGATATCCACACCCCTGCCAGAT
P P T S Q A S E G A S S D I H T P A Q M 140

961

GCTCCTGTCCACCTTGCACTCCACTCAGCGCCCCACCCCTCCCAGTAGGATCACTGTCCTC
L L S T L Q S T Q R P T L P V G S L S S 160

1021

GGACAAAGAGCTCACTCGTCCCAATGAAACCACCATTCACACTGCGGGCCACAGTCTGGC
D K E L T R P N E T T I H T A G H S L A 180

1081

AGTTGGGCCTGAGGCTGGGGAGAACCAGAAGCAGCCGGAATAATGCTGGTCCCACAGC
V G P E A G E N Q K Q P E K N A G P T A 200

1141

CAGGACATCAGCCACAGTGCCGGTCCTGTGCCTCCTGGCCATCATCTTCATCCTCACCGC
R T S A T V P V L C L L A I I F I L T A 220

1201

AGCCCTTTCTATGTGCTGTGCAAGAGGAGGGGGCAGTCACCGCAGTCTCTCCAGA
A L S Y V L C K R R R G Q S P Q S S P D 240

1261

TCTGCCGGTTCATTATATACCTGTGGCACCTGACTCTAATACCTGAGCCAAGAATGGAAG
L P V H Y I P V A P D S N T * 254

1321

CTTGTGAGGAGACGGACTCTATGTTGCCAGGCTGTTATGGAACCTGAGTCAAGTGAT

1381

CCTCCACCTTGGCCTCTGAAGGTGCGAGGATTATAGGCGTCACCTACCACATCCAGCCT

1441

ACACGTATTTGTTAATATCTAACATAGGACTAACCAGCCACTGCCCTCTCTTAGGCCCCCT

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FIGURE 1C

1501

CATT TAAA AACGGT TATA CTATA AAAATCTGCTTTT CACACTGGGTGATAATAACTTGGAC

1561

AAATCTATGTGTATTTTGTTTTGTTTTGCTTTGCTTTGTTTTGAGACGGAGTCTCGCTC

1621

TGTCATCCAGGCTGGAGTGCAGTGGCATGATCTCGGCTCACTGCAACCCCATCTCCAG

1681

GTTCAAGCGATTCTCCTGCCTCCTCCTAAGTAGCTGGGACTACAGGTGCTCACCACCACA

1741

CCCGGCTAATTTTTTGATTTTTTAGTAGAGACGGGGTTTCACCATGTTGACCAGGCTGGT

1801

CTCGAATCCTGACCTGGTGATCTGCCACCCAGGCCTCCCAAAGTGCTGGGATTAAAGG

1861

TGTGAGCCACCATGCCTGGCCCTATGTGTGTTTTTAACTACTAAAAATTATTTTTGTAA

1921

TGATTGAGTCTTCTTTATGGAAACAAC TGGCCTCAGCCCTTGCGCCCTTACTGTGATTCC

1981

TGGCTTCATTTTTTGCTGATGGTTCCCCCTCGTCCCAAATCTCTCTCCAGTACACCAGT

2041

TGTTCTCCCCACCTCAGCCCTCTCTGCATCTCTGTACCCGCAACGAAGGCCTGGG

2101

CTTTCCACCTCCCTCCTTAGCAGGTGCCGTGCTGGGACACCATACGGGTGGTTTCAC

2161

CTCCTCAGTCCCTTGCCCTACCCAGTGAGAGTCTGATCTTGTTTTATTGTTATTGCTTT

2221

TATTATTATTGCTTTTATTATCATTA AA ACTCTAGTTCTTGTTTTGTCTCTCAAAAAAA

2281

AAAAAAAAAAAAAAAAAAAAAAAAA

4/5

FIGURE 2

```

CKa-5  24  LRPGRVLLLLLLLLLVLTQPGNGNEGSVTGSCYCGKRISSDSPPSVQF 73
          :: . . . : ||||:          :...:|...: -:|||.|.|. |
MIP1-b  1  MKLCVSAFSLLLLV.....AAFCDSVLSA.PIGSDPPTSCCF 36

CKa-5  74  MNRLRKHLRAYHRCLYYTR.....FQLLSWSVCGGNKDPWVQELM 113
          || |.: . . | |.          :  : :|: :.:|||.|.:.
MIP1-b 37  SYTSRKIHRNFVMDYYETSSLCSQPAVVFLTKKGRQICADPSEPWNVEYV 86

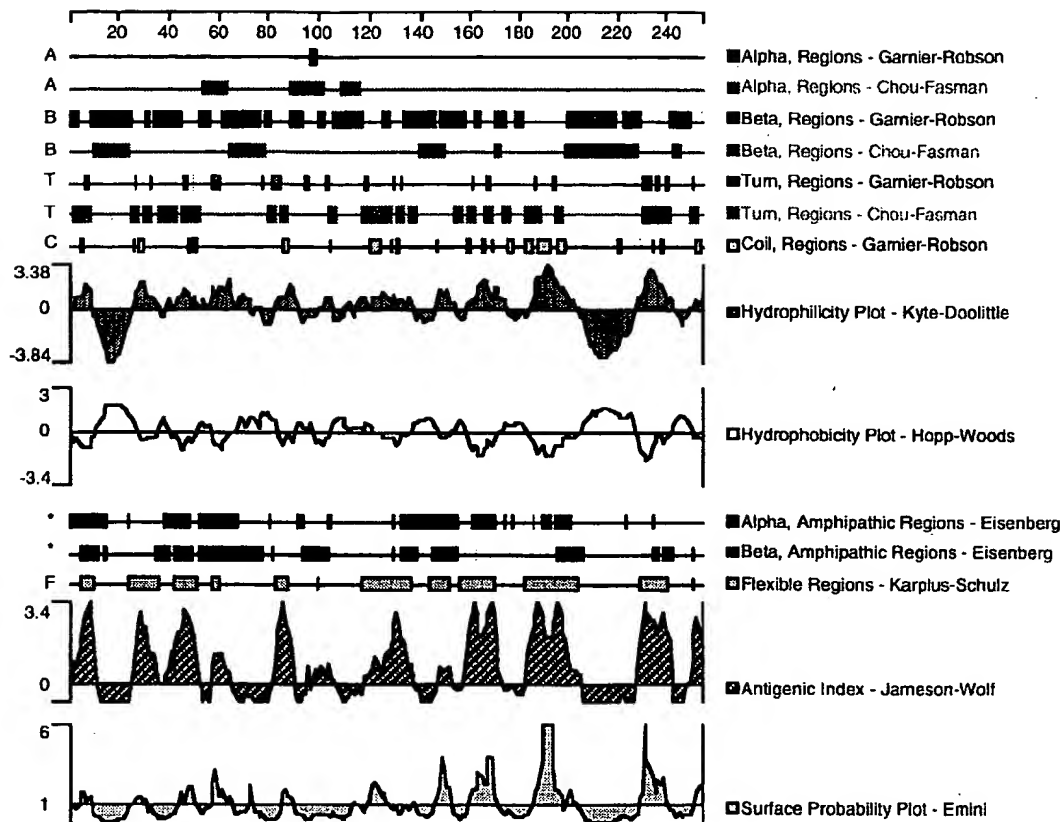
CKa-5 114  SCLDLK 119
          . |:.|.
MIP1-b 87  NDLELN 92

```

Percent Similarity: 51.220 Percent Identity: 28.049

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FIGURE 3



SEQUENCE LISTING

<110> Human Genome Sciences, Inc. et al.

<120> Chemokine Alpha-5

<130> PF401PCT

<140> Unassigned

<141> 1998-11-19

<150> 60/066,369

<151> 1997-11-21

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Applicant's or agent's file reference number	PF401PCT	International application F. MUS 98/24619
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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>2</u> , line <u>37</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input type="checkbox"/>	
Name of depositary institution American Type Culture Collection ("ATCC")	
Address of depositary institution (including postal code and country) 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	
Date of deposit 29 AUGUST 1997	Accession Number 209231
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
DNA Plasmid 2360779	
In respect to those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC).	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

For receiving Office use only	
<input type="checkbox"/> This sheet was received with the international application	Given to us by Patricia S. Schmitt IAPD - PCT Operations (703) 305-3678
Authorized officer	

For International Bureau use only	
<input type="checkbox"/> This sheet was received by the International Bureau on:	
Authorized officer	

CANADA

The applicant requests that, until either a Canadian patent has been issued ~~on the basis of~~ an application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the Commissioner of Patents only authorizes the furnishing of a sample of the deposited biological material referred to in the application to an independent expert nominated by the Commissioner, the applicant must, by a written statement, inform the International Bureau accordingly before completion of technical preparations for publication of the international application.

NORWAY

The applicant hereby requests that the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on the list of recognized experts drawn up by the Norwegian Patent Office or any person approved by the applicant in the individual case.

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Regulations), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for the international publication of the application.

Page 2

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent Office or any person by the applicant in the individual case.

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent Office or any person approved by a applicant in the individual case.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in the 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US98/24619

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350, 351; 514/2, 8, 12; 536/23.1, 23.5, 24.3, 24.31; 435/69.5, 70.1, 71.1, 71.2, 471, 325, 252.3, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

NONE

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	ROTHENBERG et al. Murine eotaxin: An eosinophil chemoattractant inducible in endothelial cells and in interleukin 4-induced tumor suppression. Proc. Natl. Acad. Sci. USA. September 1995, Vol. 92, pages 8960-8964.	1-18, 20
A	GONZALO et al. Mouse eotaxin expression parallels eosinophil accumulation during lung allergic inflammation but it is not restricted to a Th2-type response. Immunity. January 1996, Vol. 4, pages 1-14.	1-18, 20



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:

A

document defining the general state of the art which is not considered to be of particular relevance

B

earlier document published on or after the international filing date

L

document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

O

document referring to an oral disclosure, use, exhibition or other means

P

document published prior to the international filing date but later than the priority date claimed

T

later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X

document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y

document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

Z

document member of the same patent family

Date of the actual completion of the international search

03 MARCH 1999

Date of mailing of the international search report

17 MAR 1999

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

PREMA MERTZ

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/24619

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-18, 20

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US98/24619

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

C12N 5/10, 15/12, 15/19, 15/63, 15/64; A61K 38/16, 38/19; C07K 14/00, 14/47, 14/52

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

530/350, 351; 514/2, 8, 12; 536/23.1, 23.5, 24.3, 24.31; 435/69.5, 70.1, 71.1, 71.2, 471, 325, 252.3, 320.1

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, CAS ONLINE, MEDLINE, BIOSIS, CAPLUS

search terms: lymphokine, chemokine, polynucleotide, nucleic acid, chemokine alpha-5, protein, polypeptide, peptide, produce, method, treatment, therapy, administer

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claims 1-18, 20, drawn to a polynucleotide encoding chemokine alpha-5, a method for making a recombinant vector, a recombinant vector, a method of making a recombinant host cell, a recombinant host cell, a method for producing chemokine alpha-5 polypeptide, a chemokine alpha-5 polypeptide, and a method of treatment with the chemokine alpha-5 polypeptide.

Group II, claim 19, drawn to an antibody that binds to the chemokine alpha-5 polypeptide.

The inventions listed as Groups I-II do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

Pursuant to 37 C.F.R. § 1.475 (d), the ISA/US considers that where multiple products and processes are claimed, the main invention shall consist of the first invention of the category first mentioned in the claims and the first recited invention of each of the other categories related thereto. Accordingly, the main invention (Group I) comprises the first-recited product, a polynucleotide encoding chemokine alpha-5, a method for making a recombinant vector, a recombinant vector, a method of making a recombinant host cell, a recombinant host cell, a method for producing chemokine alpha-5 polypeptide, a chemokine alpha-5 polypeptide, and a method of treatment with the chemokine alpha-5 polypeptide. Further pursuant to 37 C.F.R. § 1.475 (d), the ISA/US considers that any feature which the subsequently recited products and methods share with the main invention does not constitute a special technical feature within the meaning of PCT Rule 13.2 and that each of such products and methods accordingly defines a separate invention.